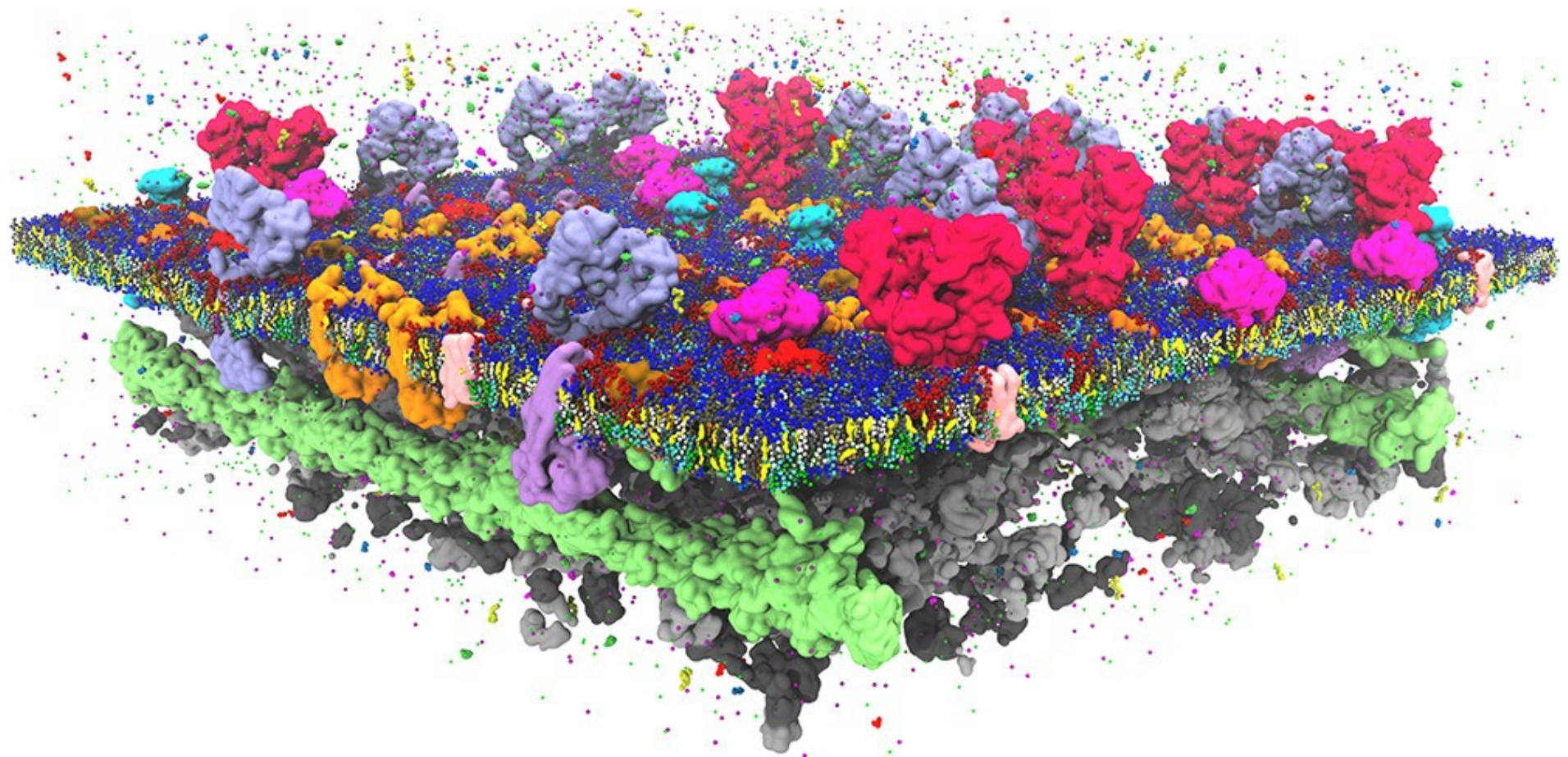


BC 03 :

Membranes et échanges membranaires



Introduction

- Lipides amphiphiles
- Liaisons de Van der Waals
- Différents rôles

FIGURE 1.5 Organization of amphipathic lipid molecules in a bilayer.

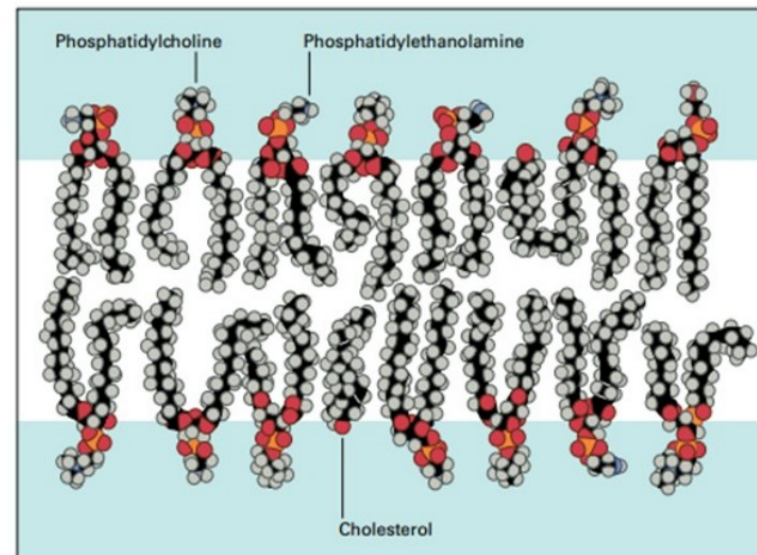
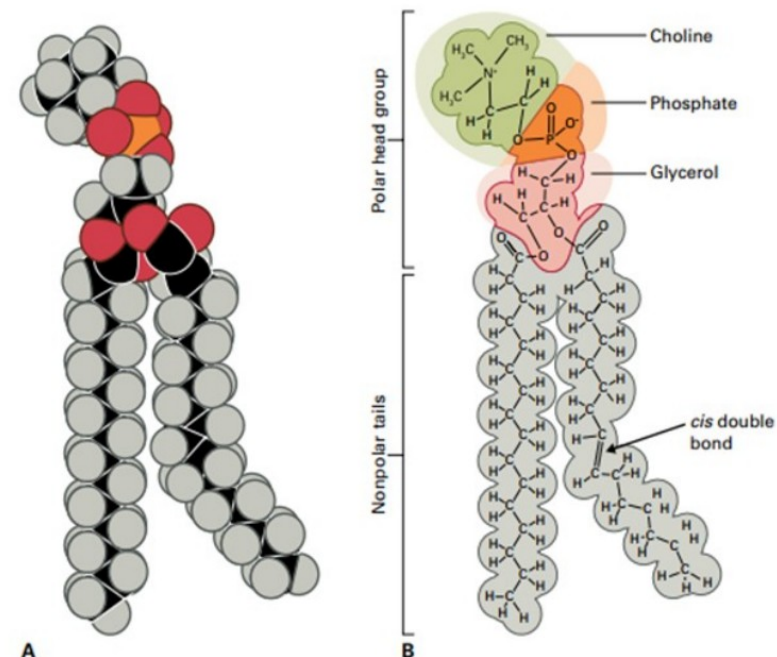
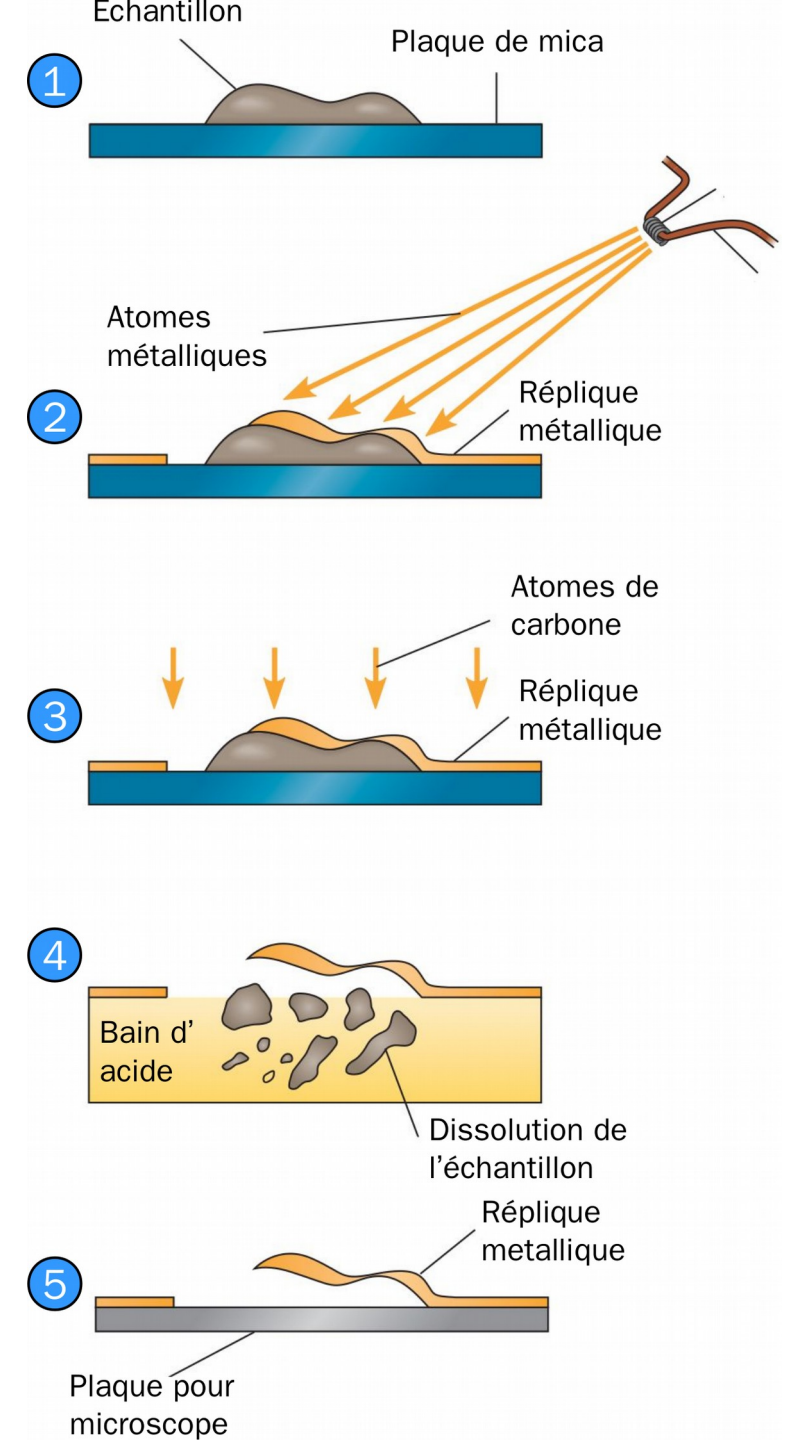
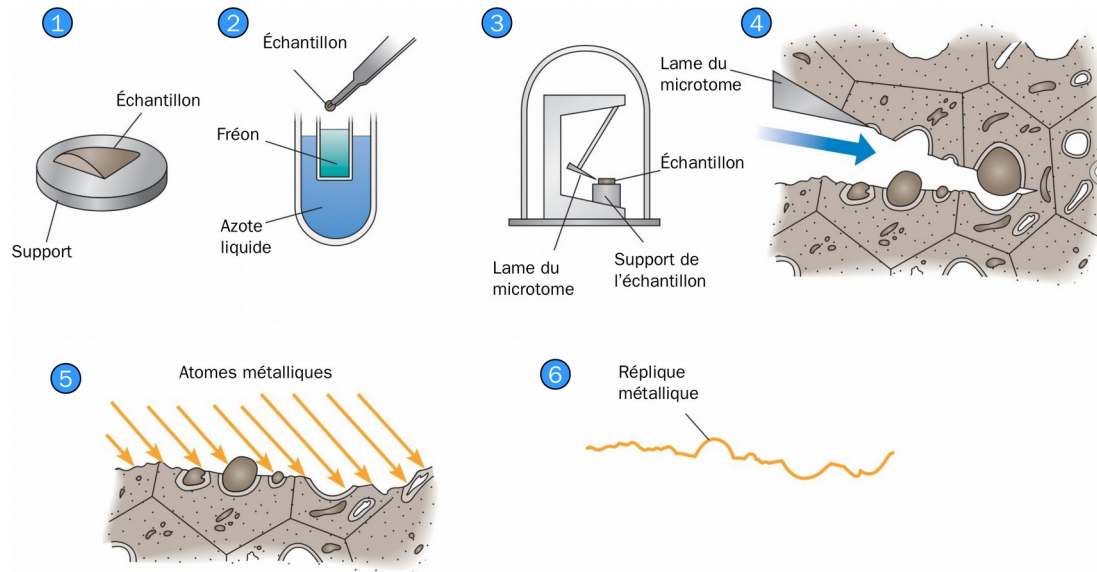


FIGURE 1.6 (A) Space-filling model of a phosphatidylcholine molecule. (B) Diagram defining the functional groups of a phosphatidylcholine molecule.



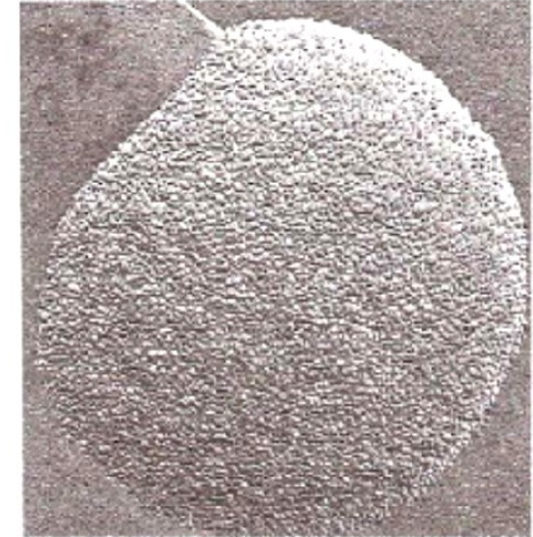
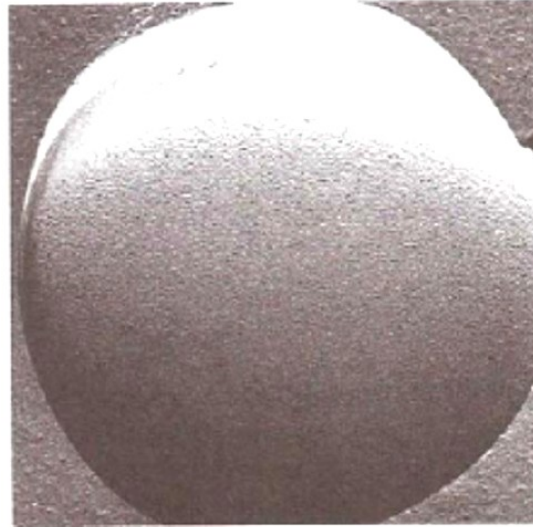
Singer et Nicholson

- Cryofracture
- Cryodécapage
- Ombrage métallique



Singer et Nicholson

- Cryofracture
- Cryodécapage
- Ombrage métallique



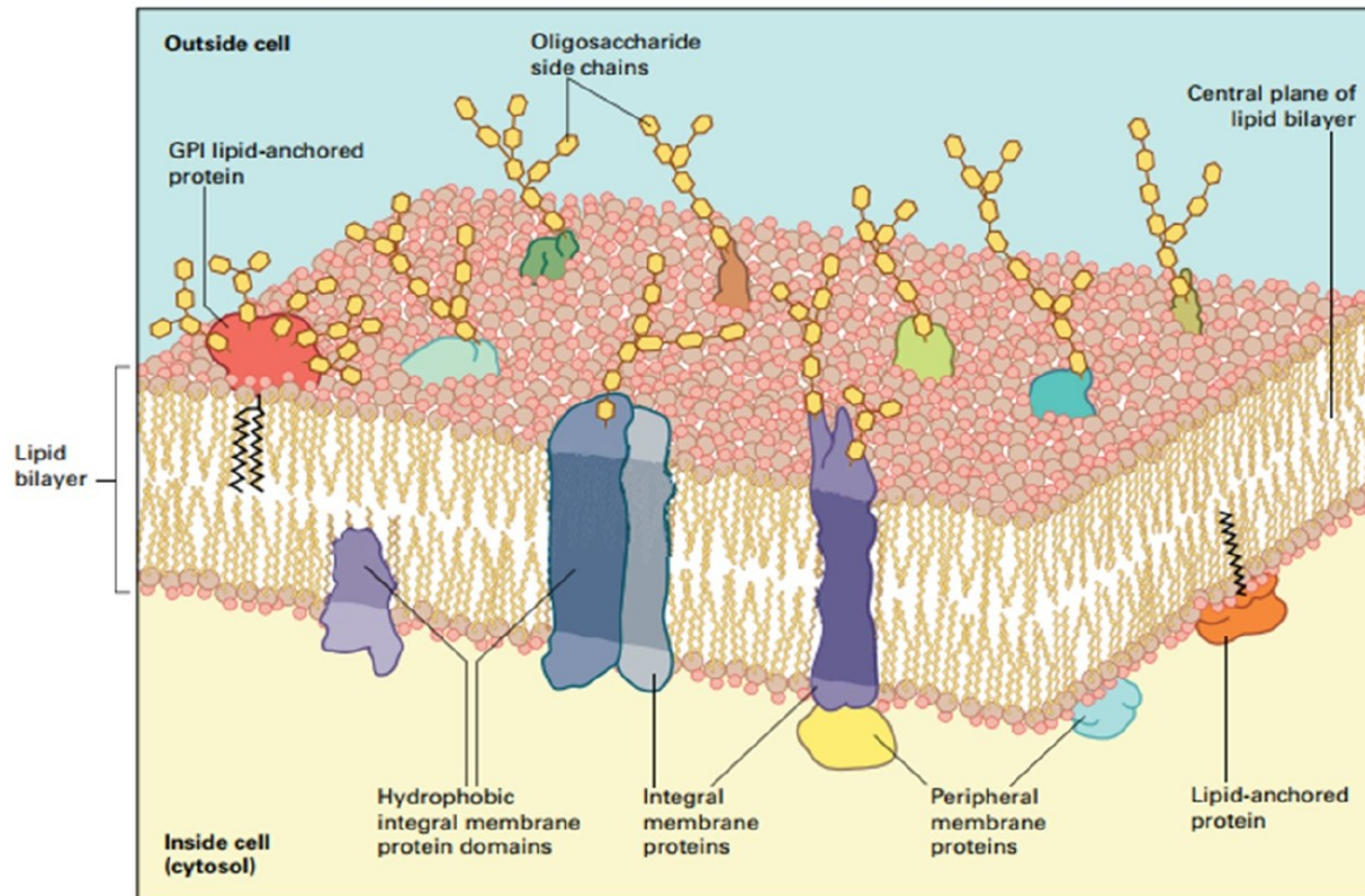
Document 1 : Clichés de cryofracture au microscope électronique d'un liposome et d'une membrane. A gauche, cryofracture de liposomes (agrandissement : 80000). A droite, cryofracture d'une membrane cytoplasmique d'E. coli (agrandissement : 50000) ; Les irrégularités visibles sur le cliché de droite disparaissent après un traitement aux protéases.

D'après Biochimie et biophysique des membranes, Shechter E., 1997, Masson, 2^e édition.

DOC 3 : Résultats d'expériences de cryofracture sur liposome ou sur membrane

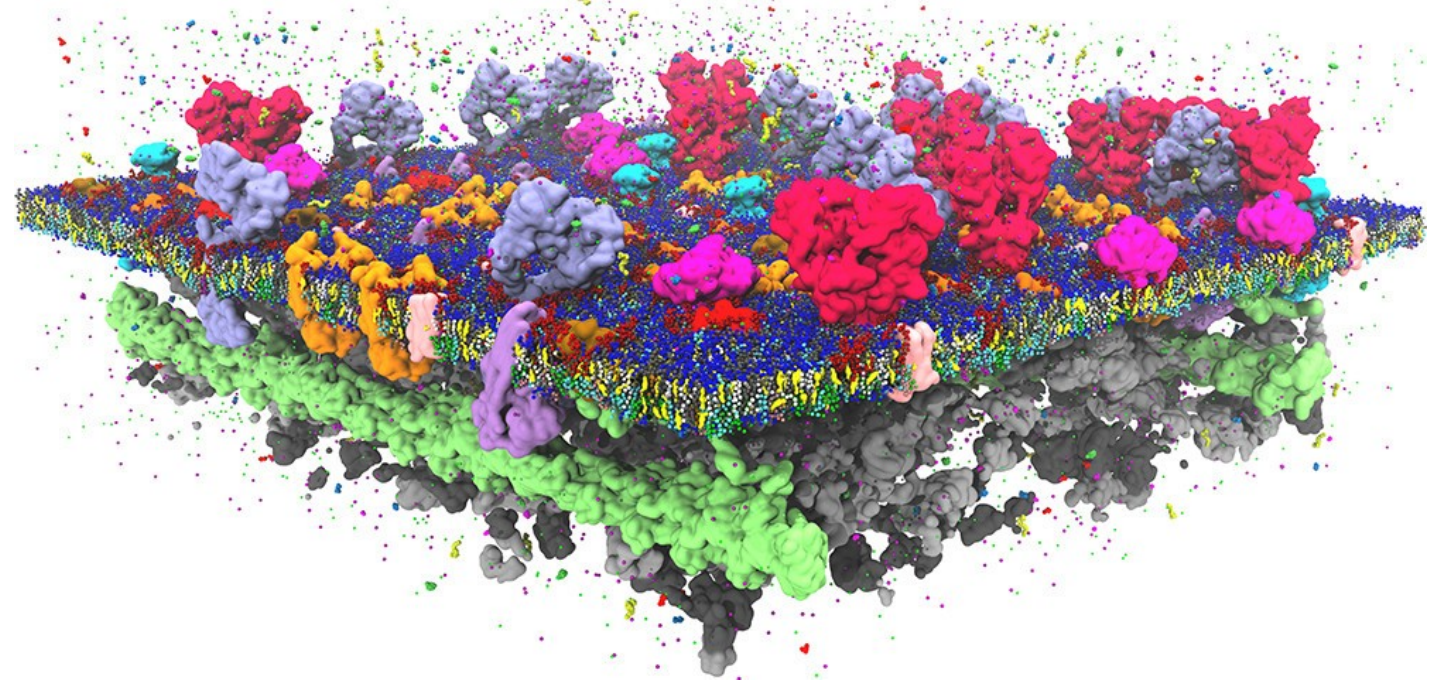
Singer et Nicholson

- Modèle final valant un prix Nobel



Protéines membranaires

- Intrinsèques/intégrales
- Extrinsèques
- Rôle des protéines
 - Adhérence
 - Marqueurs
 - Récepteurs
 - Canaux
 - Pompes (coûte de l'énergie sous forme d'ATP)
 - Transporteurs
- 60 % des médicaments ciblent des protéines membranaires



**CHEMICAL
REVIEWS**

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Review

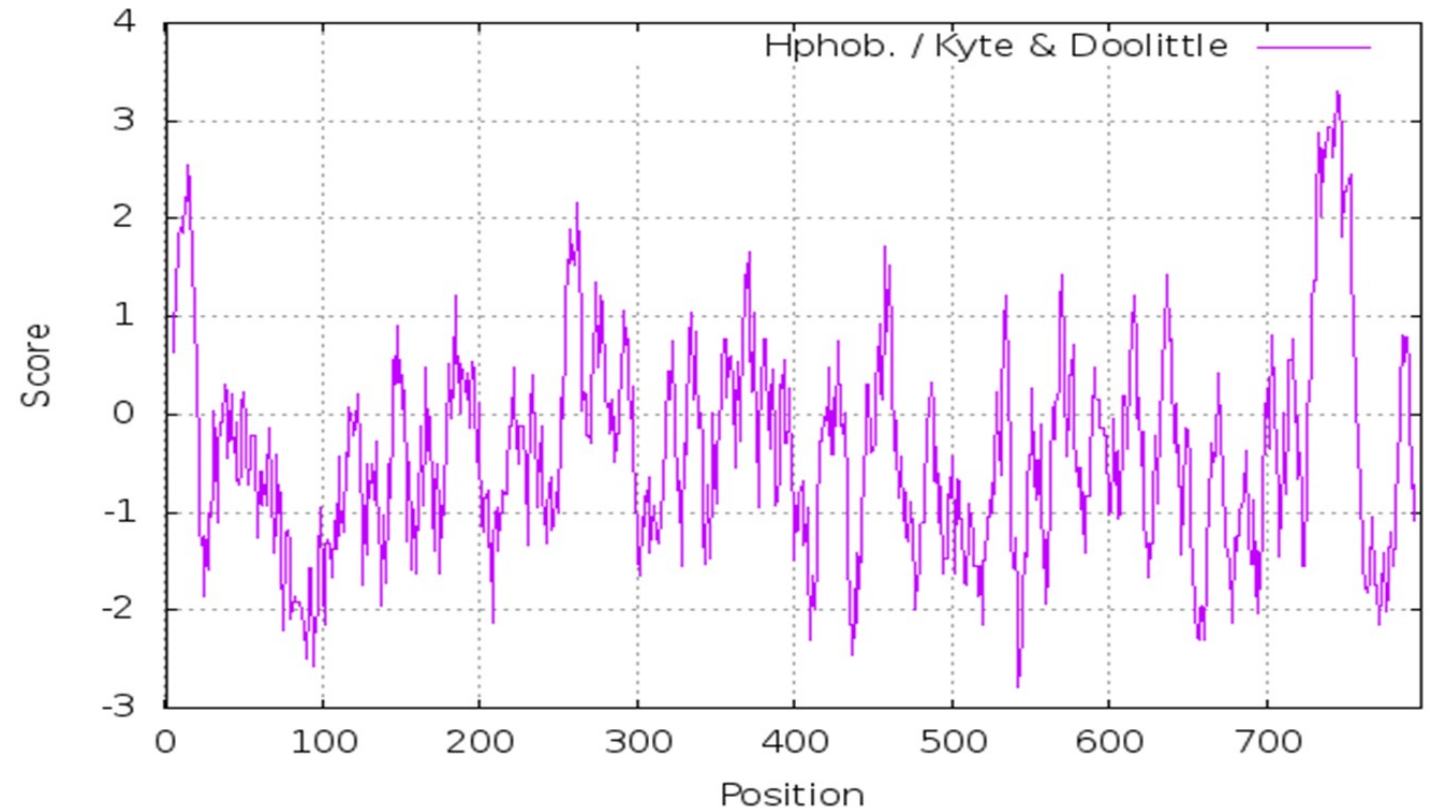
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Computational Modeling of Realistic Cell Membranes

Siewert J. Marrink,^{*,†} Valentina Corradi,[‡] Paulo C.T. Souza,[†] Helgi I. Ingólfsson,[§] D. Peter Tieleman,[‡] and Mark S.P. Sansom^{||}

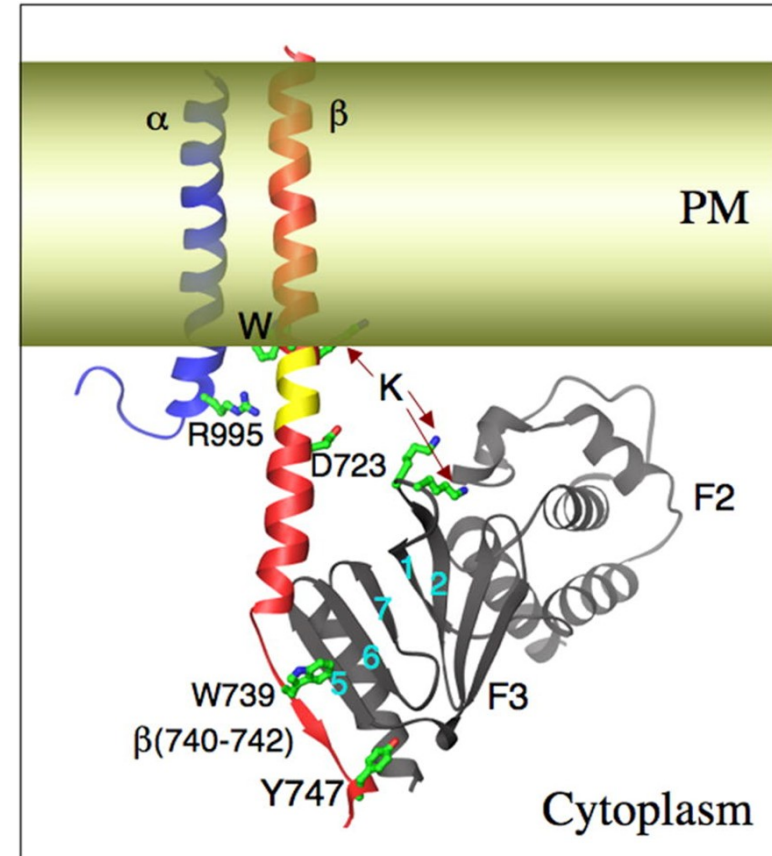
Mise en évidence d'une protéine intrinsèque

- Profil d'hydrophathie
- Ici exemple d'une protéine appelée « intégrine beta »



Mise en évidence d'une protéine intrinsèque

- Une longue portion transmembranaire constituée d'acides aminés hydrophobes



Asymétrie des membranes

- Un glycocalyx (revêtement glucidique) à l'extérieur
- Ici sur la membrane d'une hématie

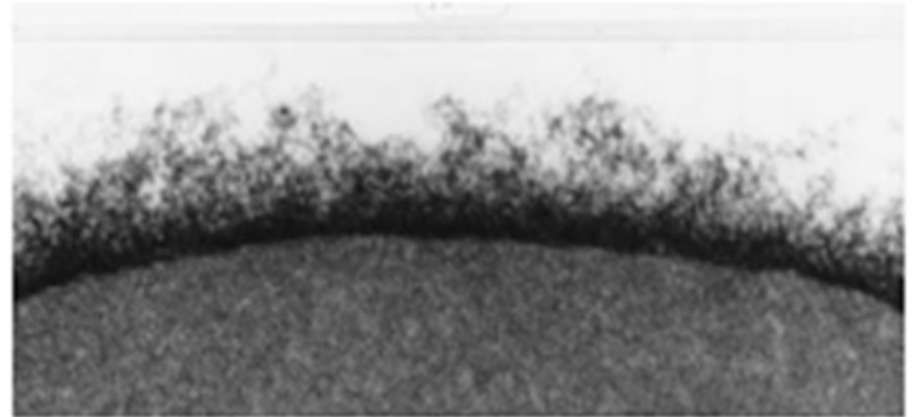
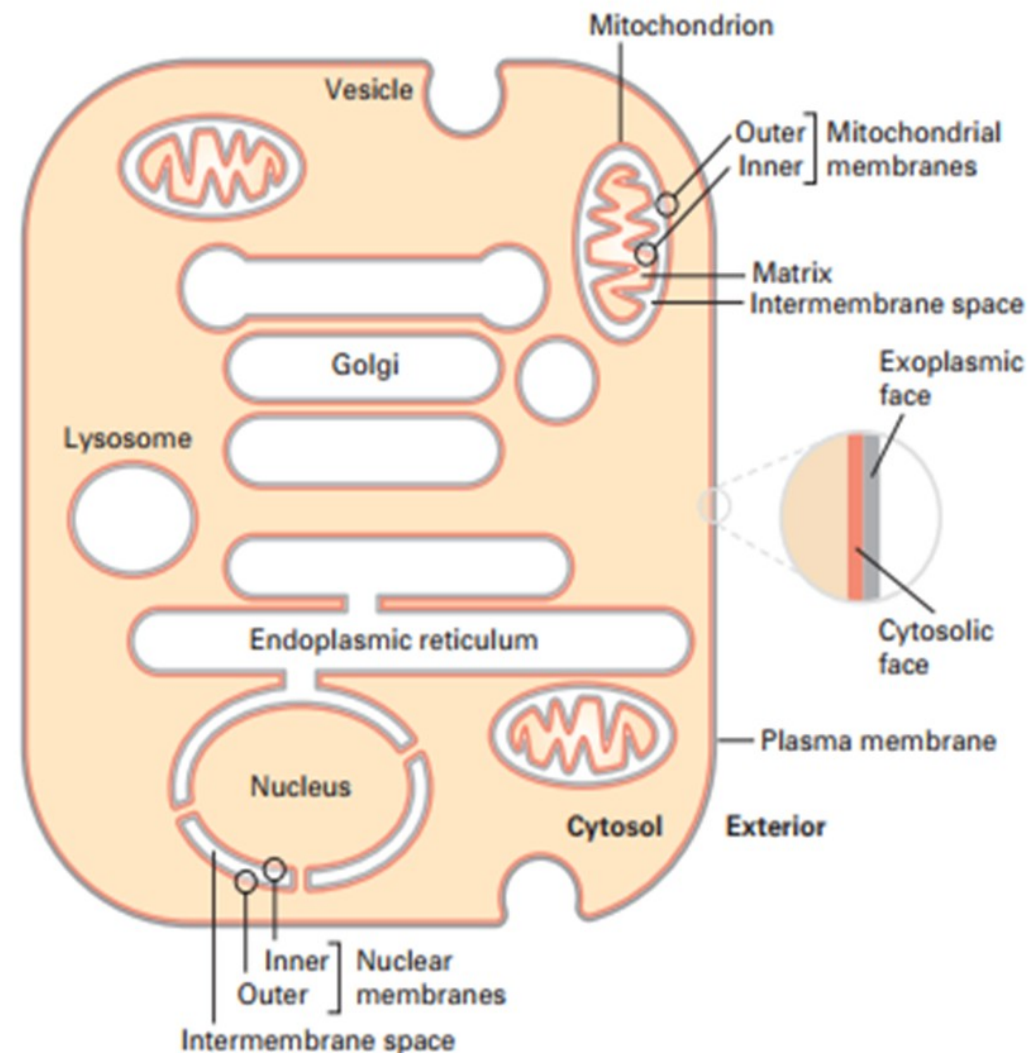


Figure 12-40 The erythrocyte glycocalyx as revealed by electron microscopy using special staining techniques. It is up to 1400 Å thick and composed of closely packed, 12- to 25-Å-diameter oligosaccharide filaments linked to plasma membrane-associated proteins and lipids. [Courtesy of Harrison Latta, UCLA.]

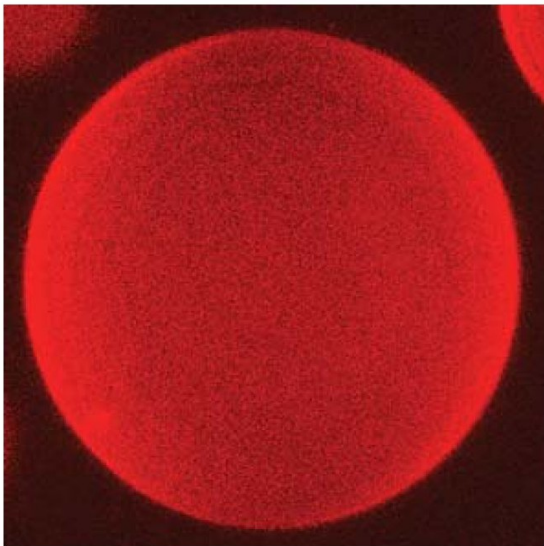
Asymétrie des membranes

- Deux hémimembranes différentes



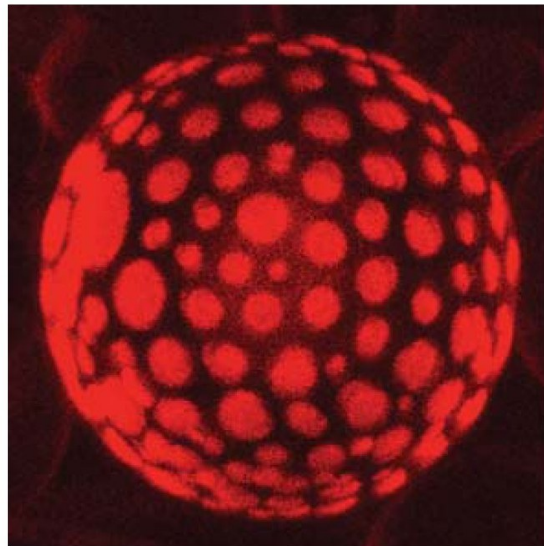
Radeaux lipidiques

- Zone moins fluide
- Sphingolipide + Cholestérol
- Caveolin



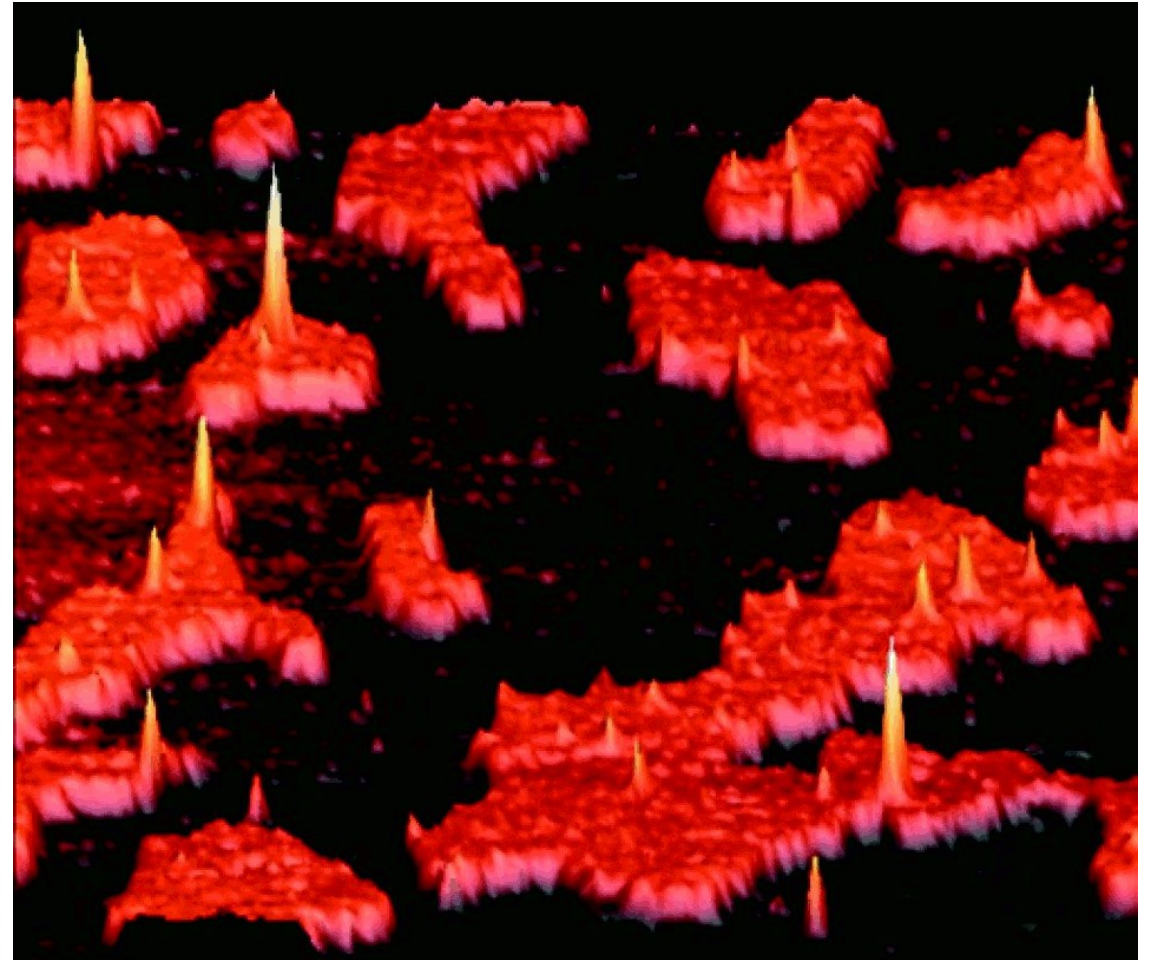
(A)

10 μm



(B)

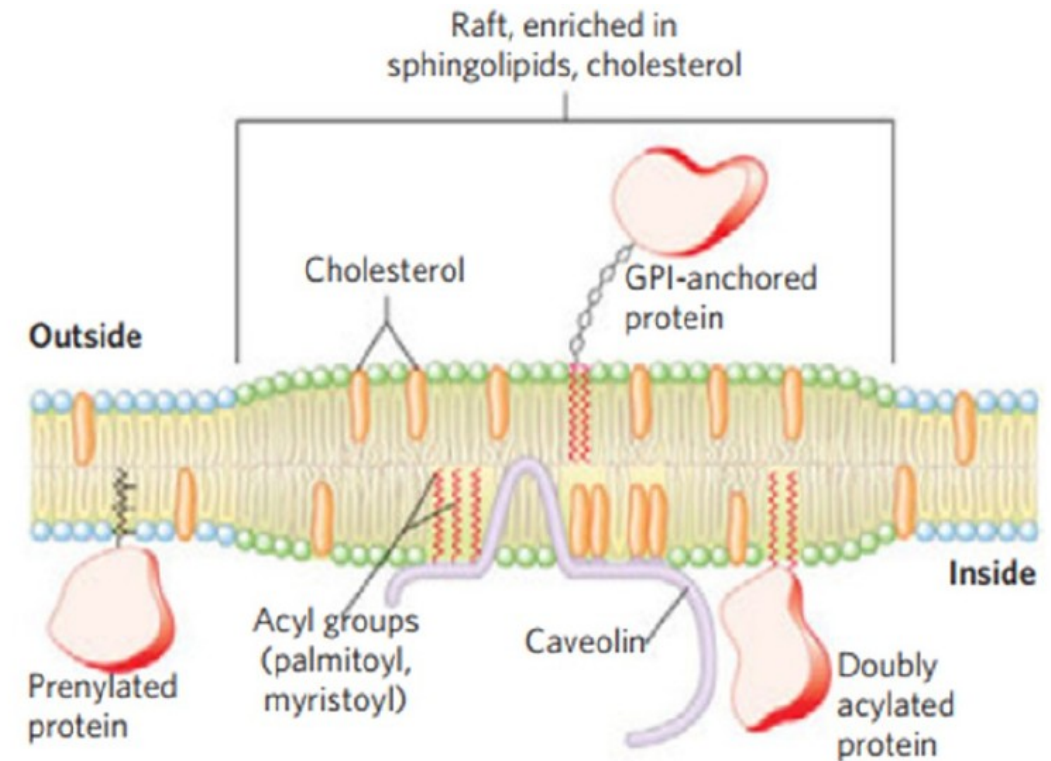
5 μm



Radeaux lipidiques

- Zone moins fluide
- Sphingolipide + Cholestérol
- Caveolin

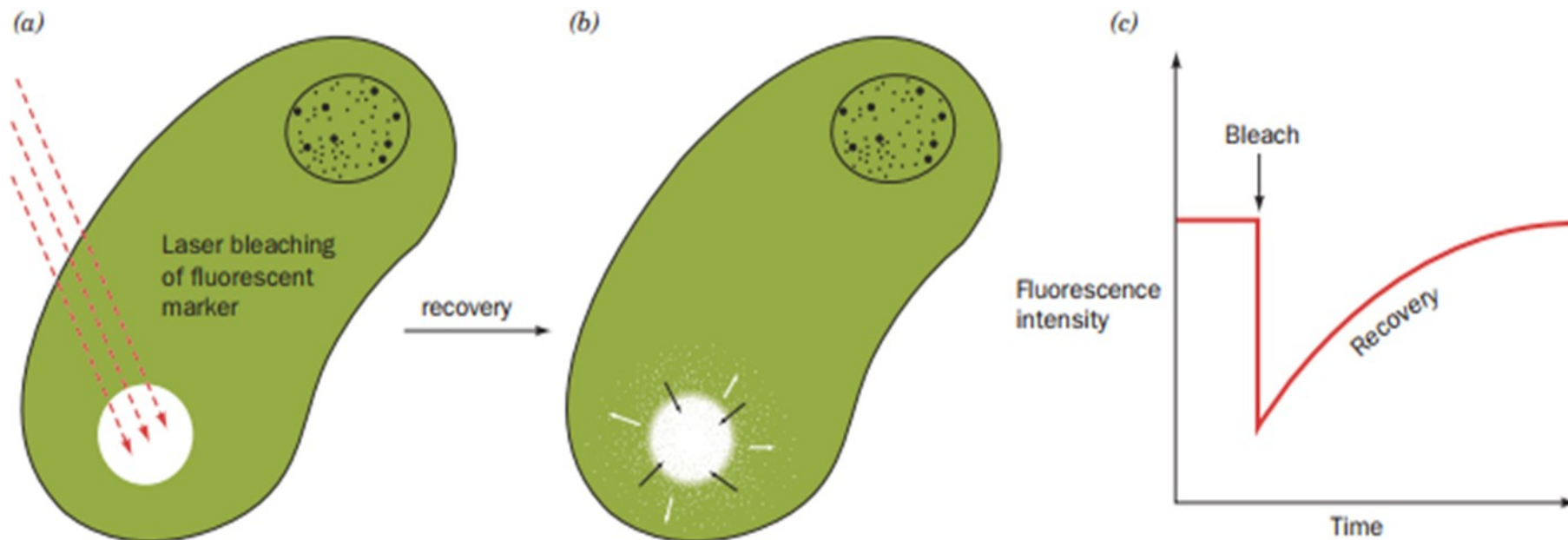
DOC 7: Organisation d'un radeau lipidique



In [molecular biology](#), **caveolins** are a family of [integral membrane proteins](#) that are the principal components of [caveolae](#) membranes and involved in receptor-independent [endocytosis](#).^{[1][2][3]} Caveolins may act as [scaffolding proteins](#) within caveolar membranes by compartmentalizing and concentrating signaling molecules. They also induce positive (inward) membrane curvature by way of oligomerization, and hairpin insertion. Various classes of signaling molecules, including [G-protein](#) subunits, receptor and non-receptor [tyrosine kinases](#), endothelial [nitric oxide synthase](#) (eNOS), and small [GTPases](#), bind Cav-1 through its 'caveolin-scaffolding domain'.

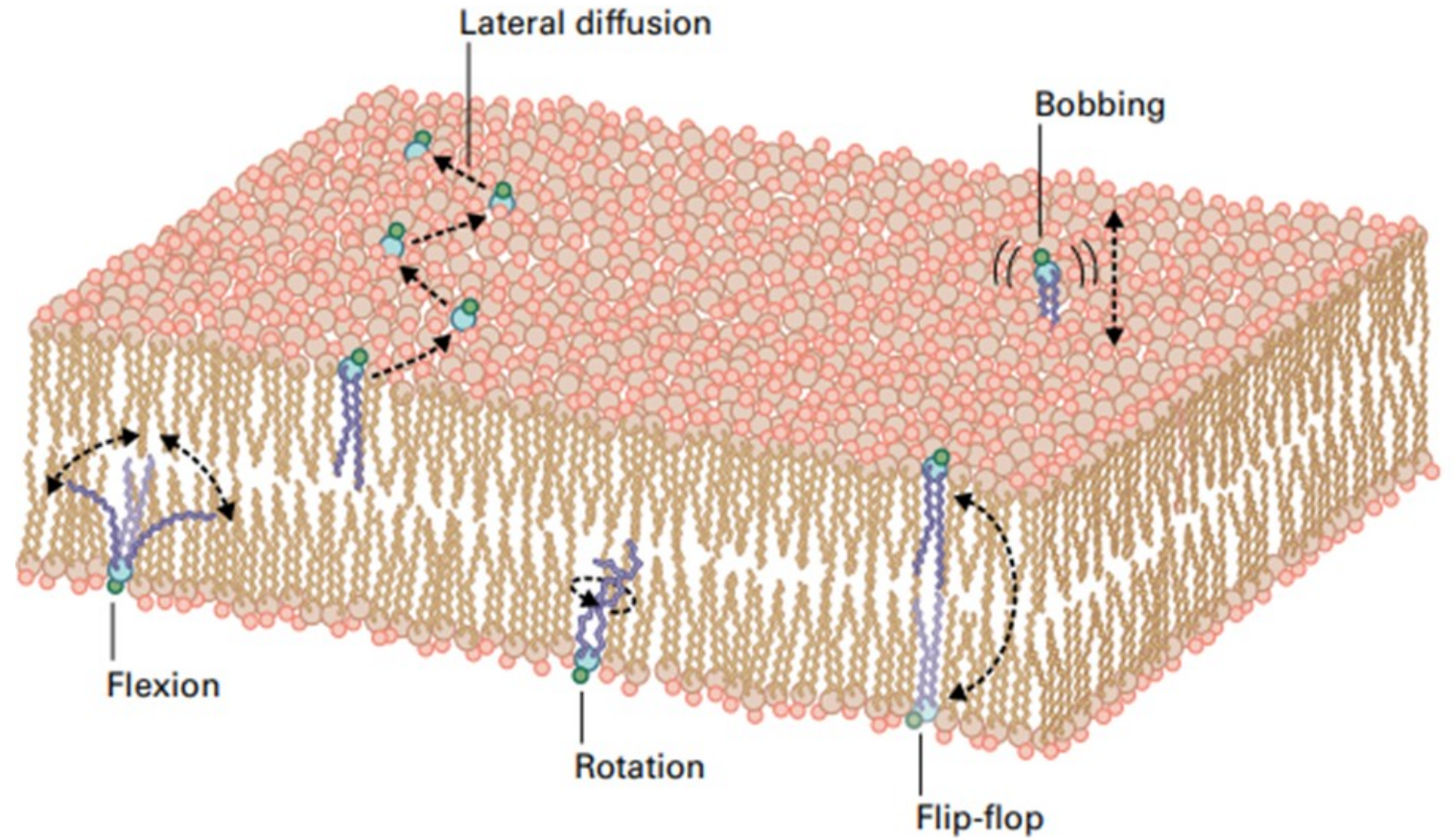
Mouvements au sein de la membrane

- Mise en évidence de la fluidité latérale par la technique de FRAP



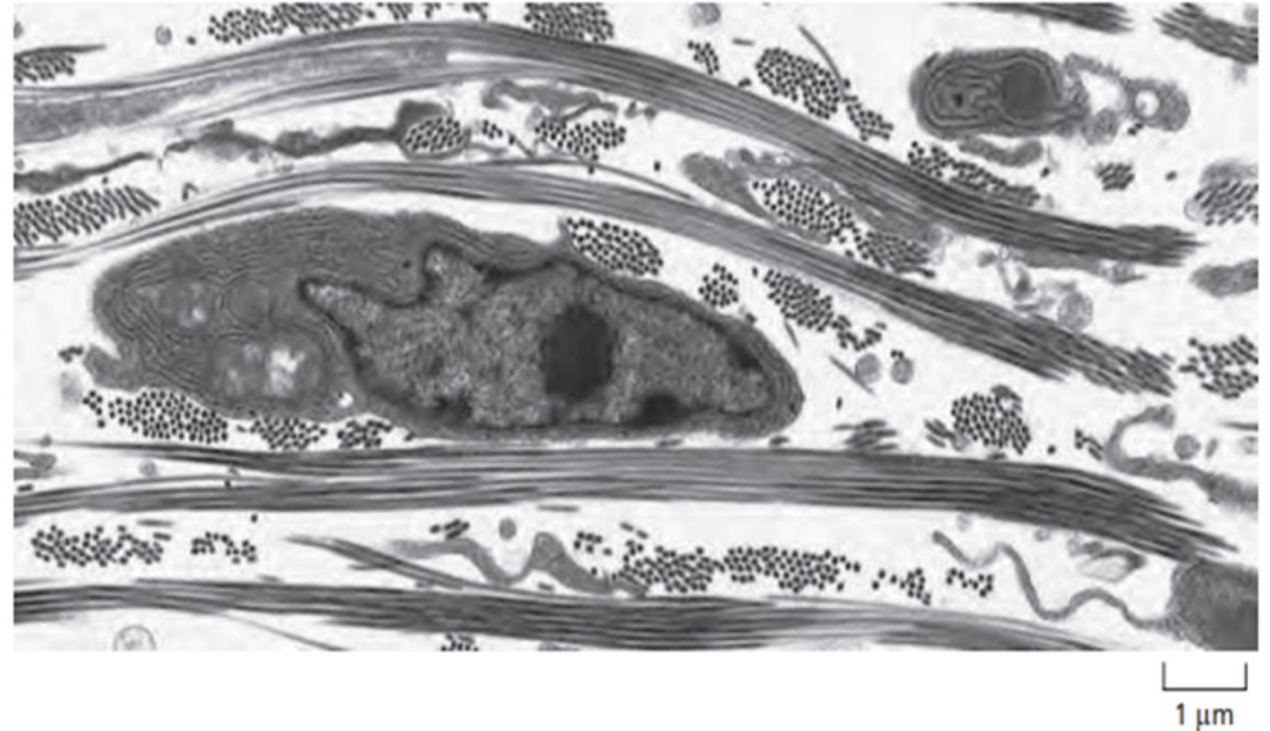
Mouvements au sein de la membrane

- Diffusion latérale
- Flip-flop
- Rotation



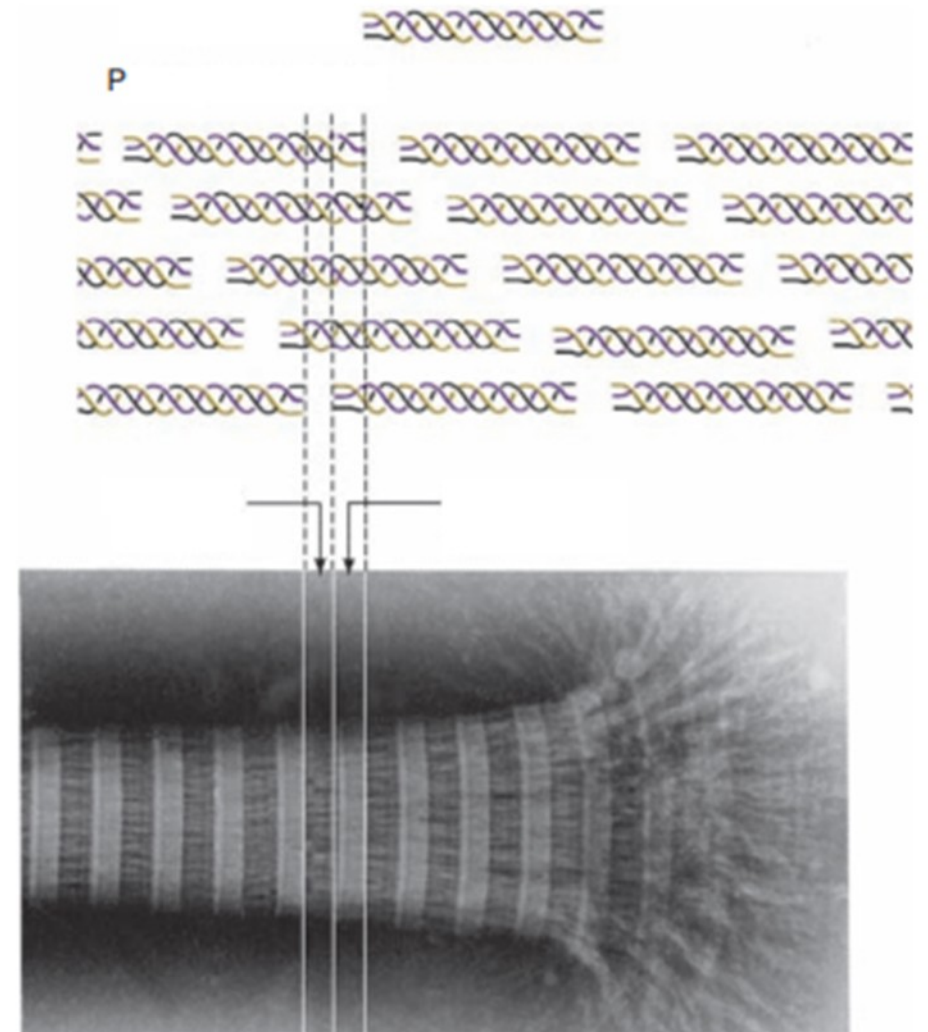
Une cellule entourée de MEC

- Exemple d'un fibroblaste entouré de fibres de collagène



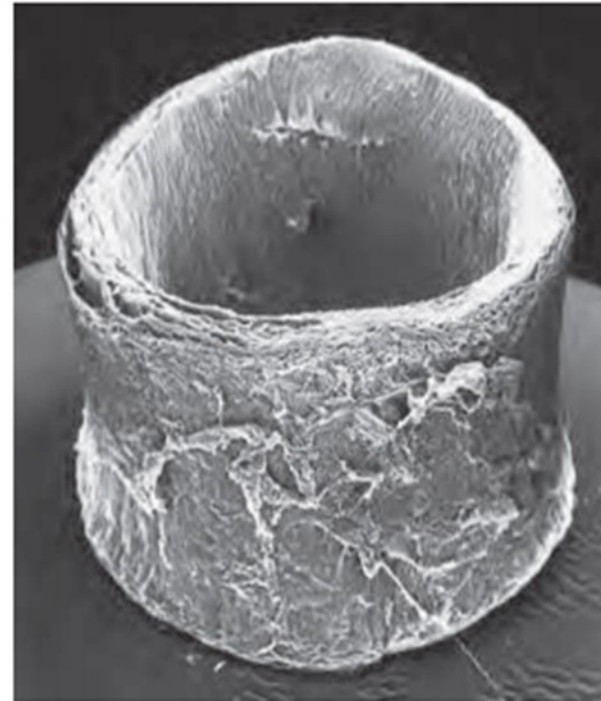
Organisation du collagène

- Stries apparaissant au MET



MEC et élastine

- Ici observées au MEB
- Abondante dans les parois des artères



(A)

1 mm



(B)

100 μm

MEC et protéoglycanes

- Grandes molécules qui retiennent l'eau dans la MEC

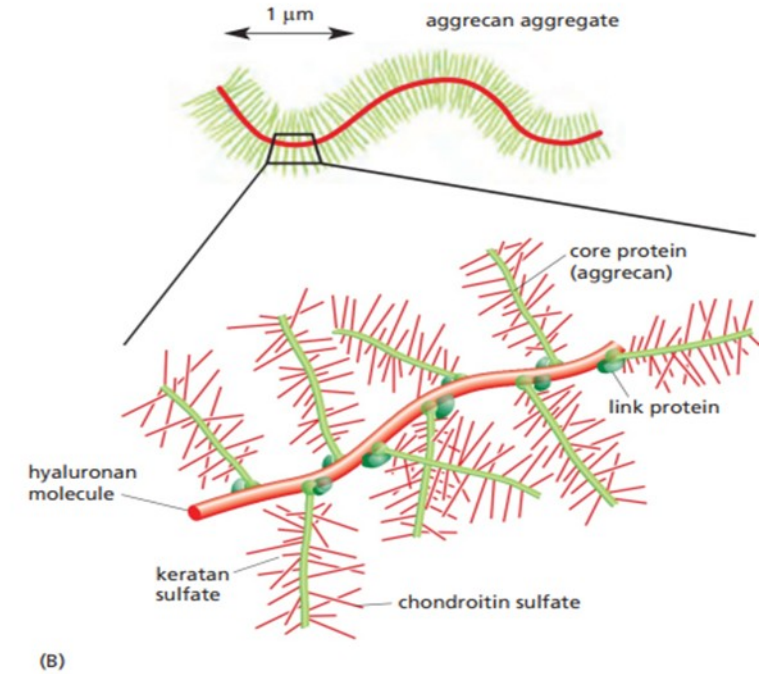
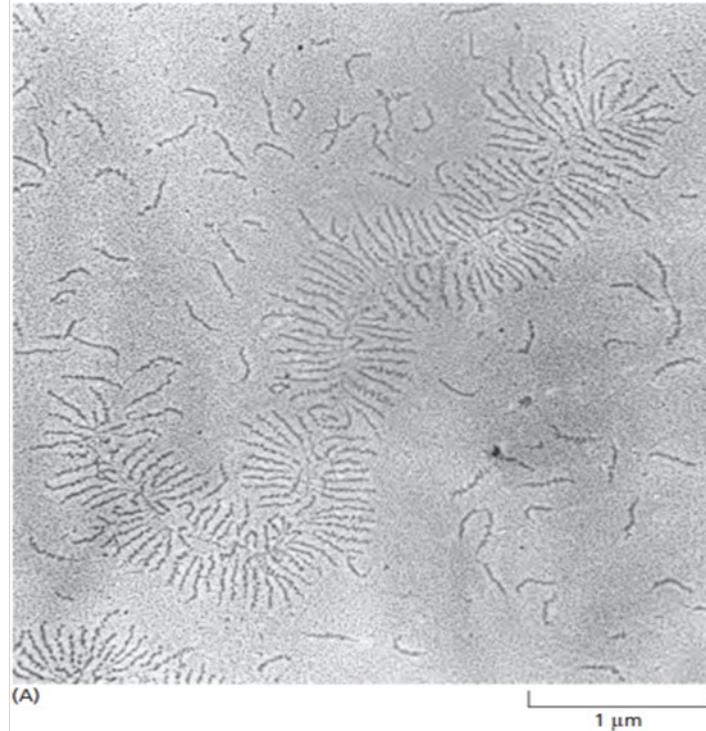


Figure 19-37 An aggrecan aggregate from fetal bovine cartilage. (A) An electron micrograph of an aggrecan aggregate shadowed with platinum. Many free aggrecan molecules are also visible. (B) A drawing of the giant aggrecan aggregate shown in (A). It consists of about 100 aggrecan monomers (each like the one shown in Figure 19-36) noncovalently bound through the N-terminal domain of the core protein to a single hyaluronan chain. A link protein binds both to the core protein of the proteoglycan and to the hyaluronan chain, thereby stabilizing the aggregate. The link proteins are members of a family of hyaluronan-binding proteins, some of which are cell-surface proteins. The molecular mass of such a complex can be 10^8 daltons or more, and it occupies a volume equivalent to that of a bacterium, which is about 2×10^{-12} cm^3 . (A, courtesy of Lawrence Rosenberg.)

MEC et GAG

- Les GAG retiennent l'eau en raison de leurs groupements chargés qui diminuent le potentiel osmotique de la MEC

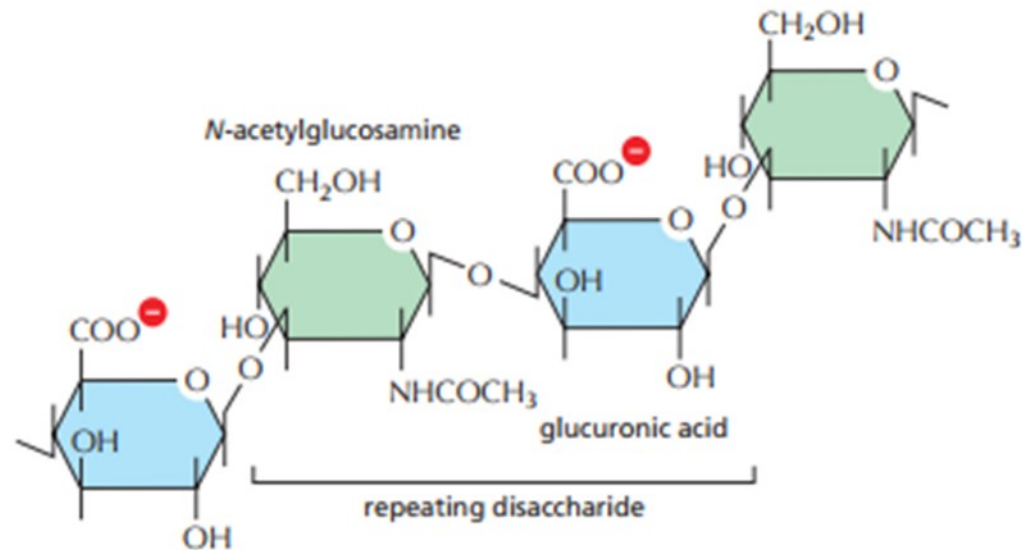
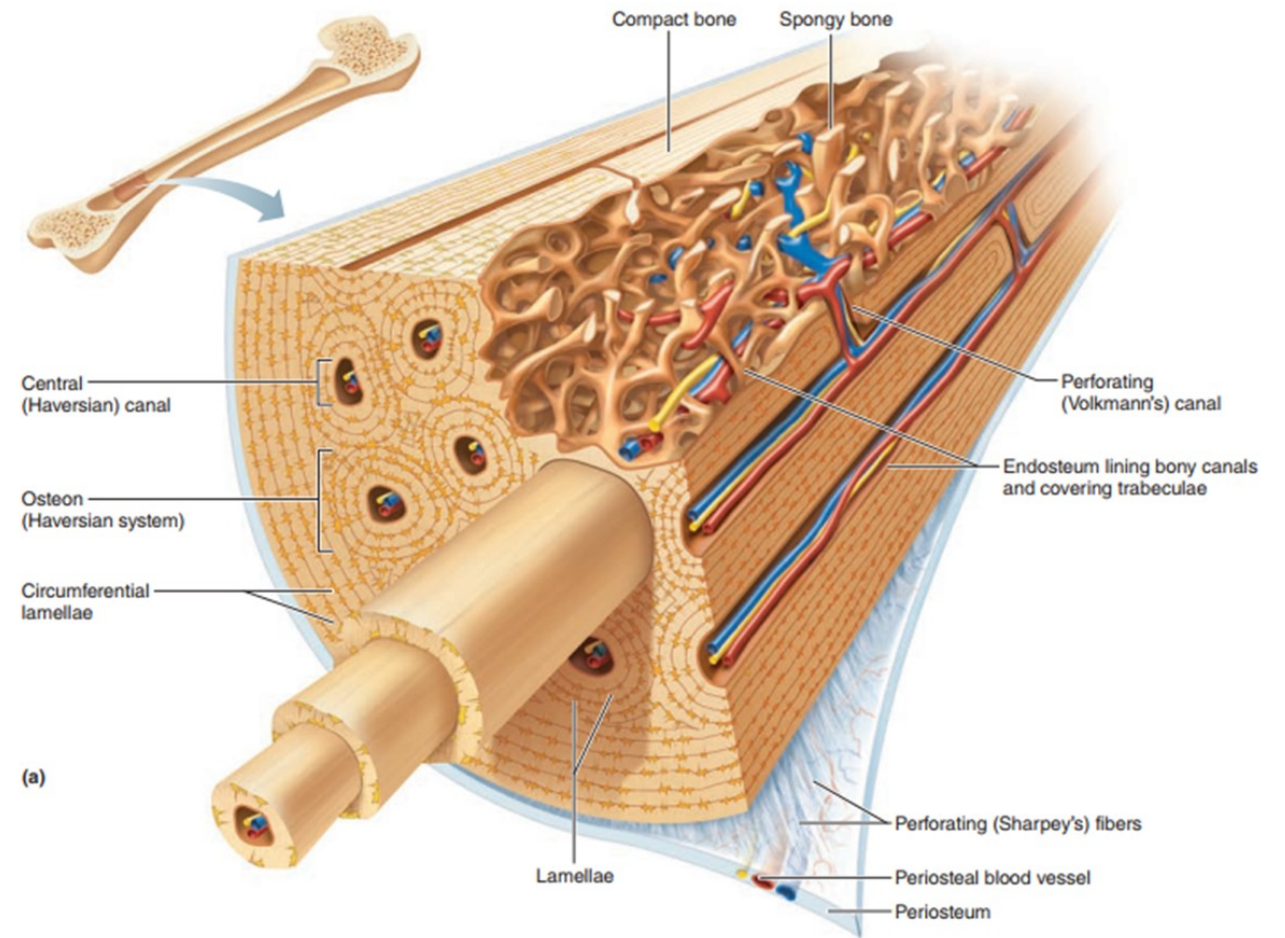


Figure 19–34 The repeating disaccharide sequence in hyaluronan, a relatively simple GAG. This ubiquitous molecule in vertebrates consists of a single long chain of up to 25,000 sugar monomers. Note the absence of sulfate groups.

Une MEC minéralisée, l'os

- Os constitué de cellules entourées d'une MEC minéralisée
- Cristaux d'hydroxyapatite (phosphate de calcium)
- Os = tissu riche en vaisseaux sanguins et nerfs



Cytosquelette

- Cellule après action d'un détergent qui solubilise les membranes
- Le cytosol contient un réseau de filaments

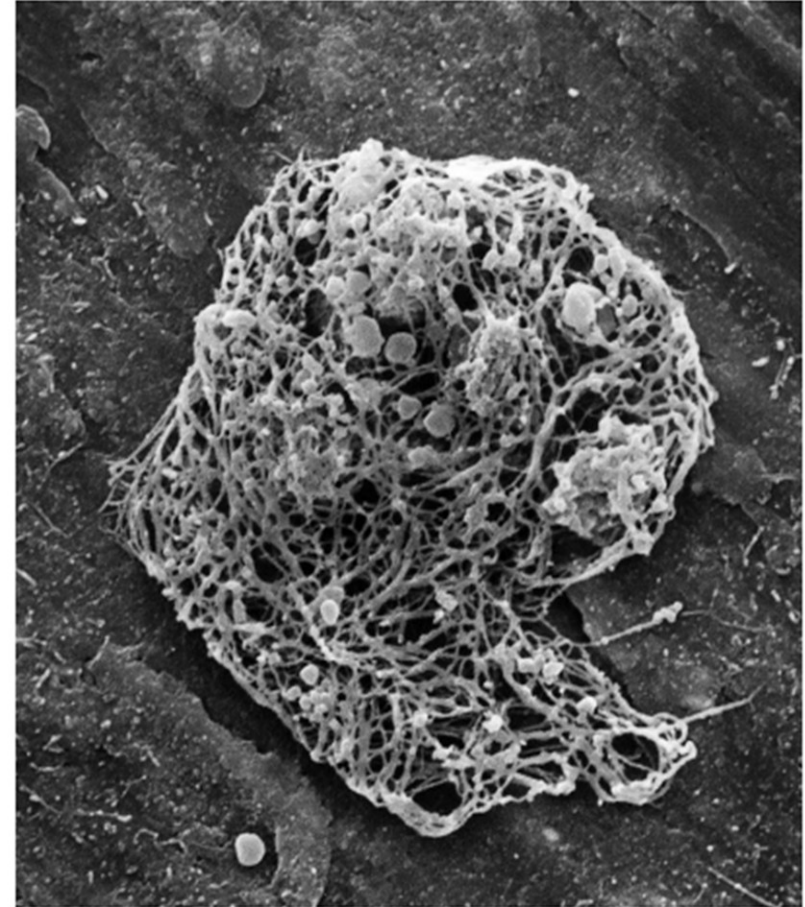
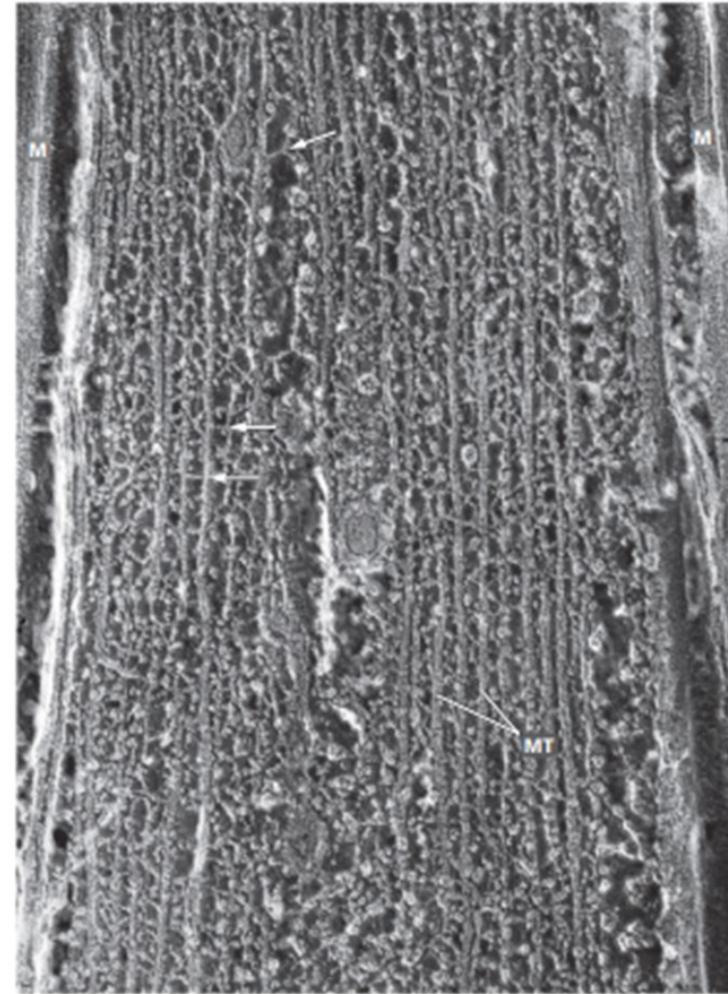


FIGURE 5.2 Electron micrograph of cytoskeletal meshwork, showing detergent-resistant fibrous elements from a suspension-cultured carrot (*Daucus carota*) cell.

Source: Xu et al. (1992). *Plant Cell* 4:941–951.

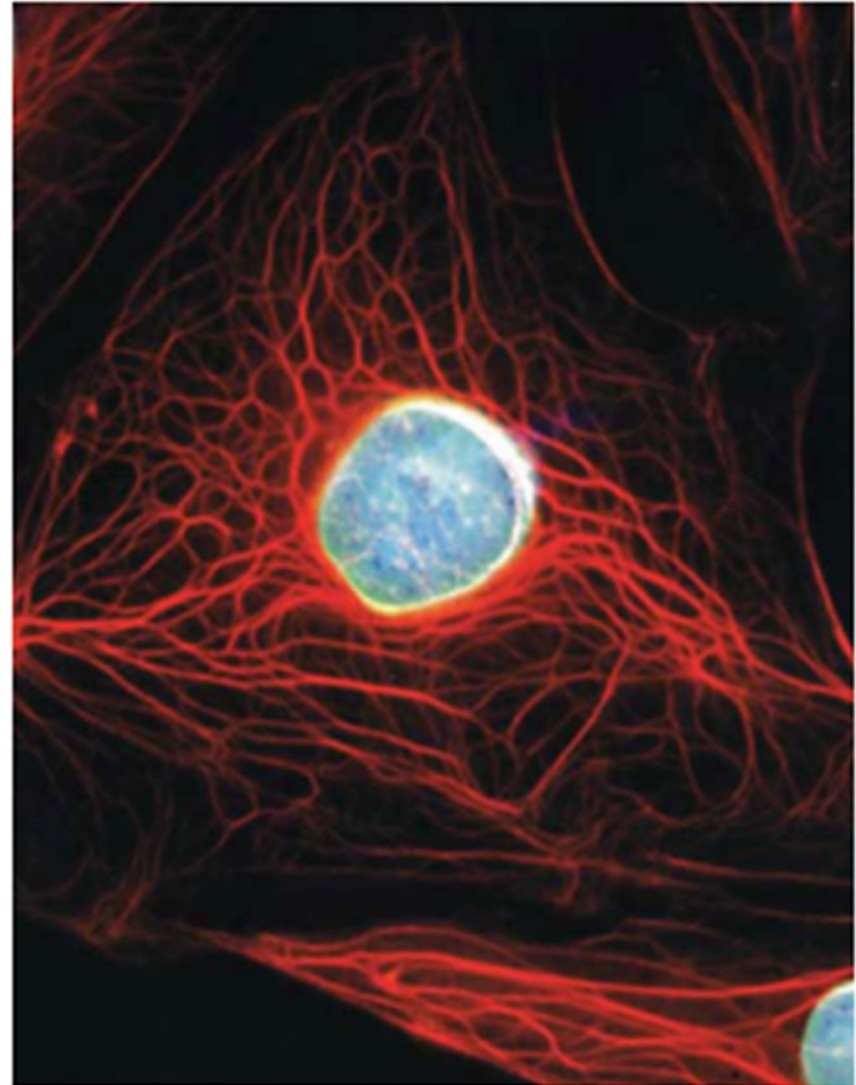
Cytosquelette : microtubules

- Axone en microscopie électronique
- Nombreux microtubules visibles



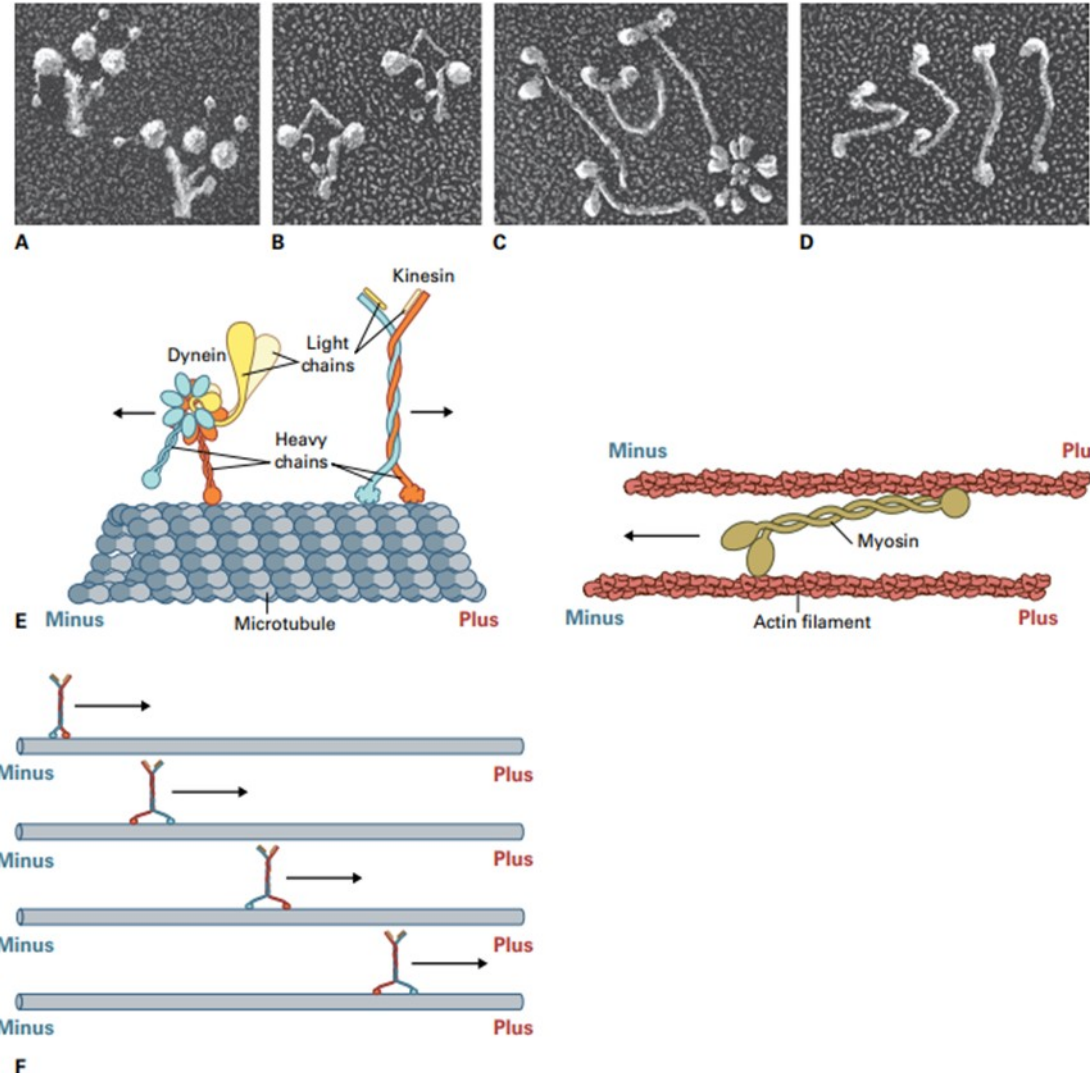
Cytosquelette : filaments intermédiaires

- Microscopie à fluorescence
- Kératine (en rouge)
- Lamines (en bleu)



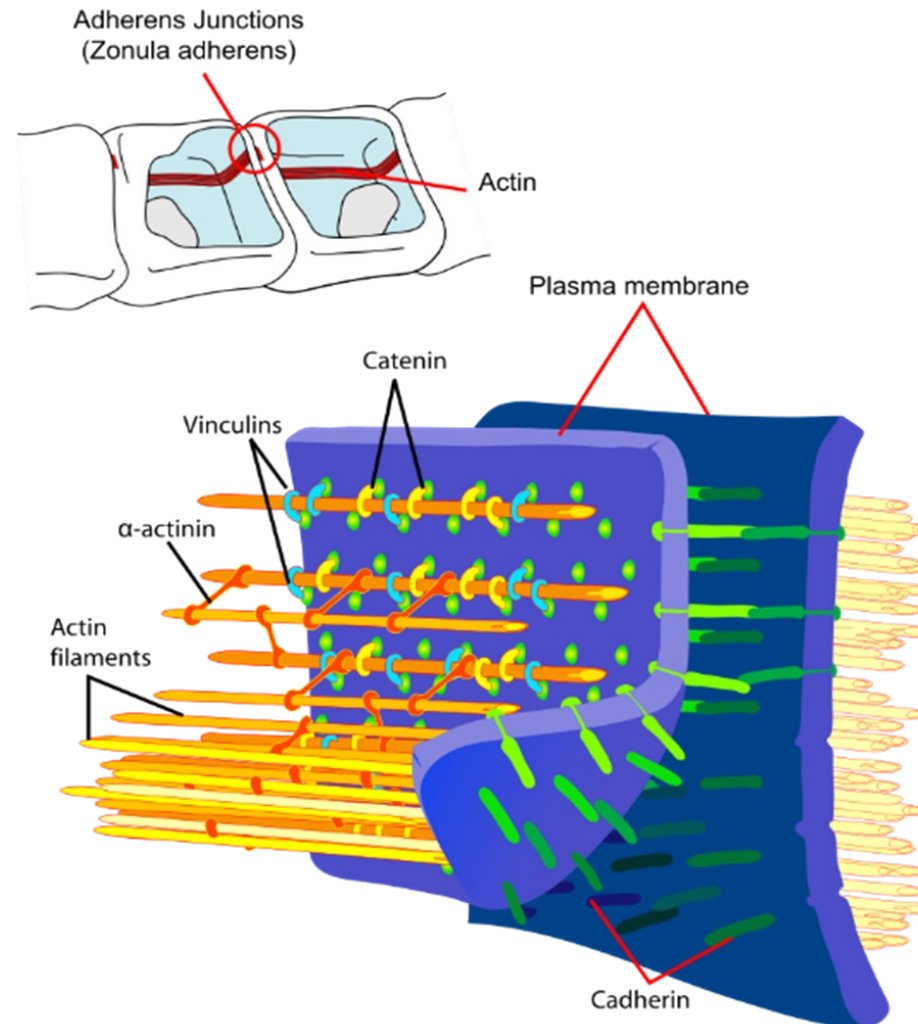
Cytosquelette : moteurs moléculaires

- Myosines et actine
- Kinésine/dynéine et microtubules
- Déplacements des organites et des protéines sur les filaments du cytosquelette



Jonctions : ceinture d'adhérence

- Pôle apical
- Dans les cellules épithéliales
- Cadhérines, actine, protéines adaptatrices



Jonctions : ceinture d'étanchéité

- Pôle apical
- Dans les cellules épithéliales
- Occludines, claudines



(a)

0.2 μm 

(b)

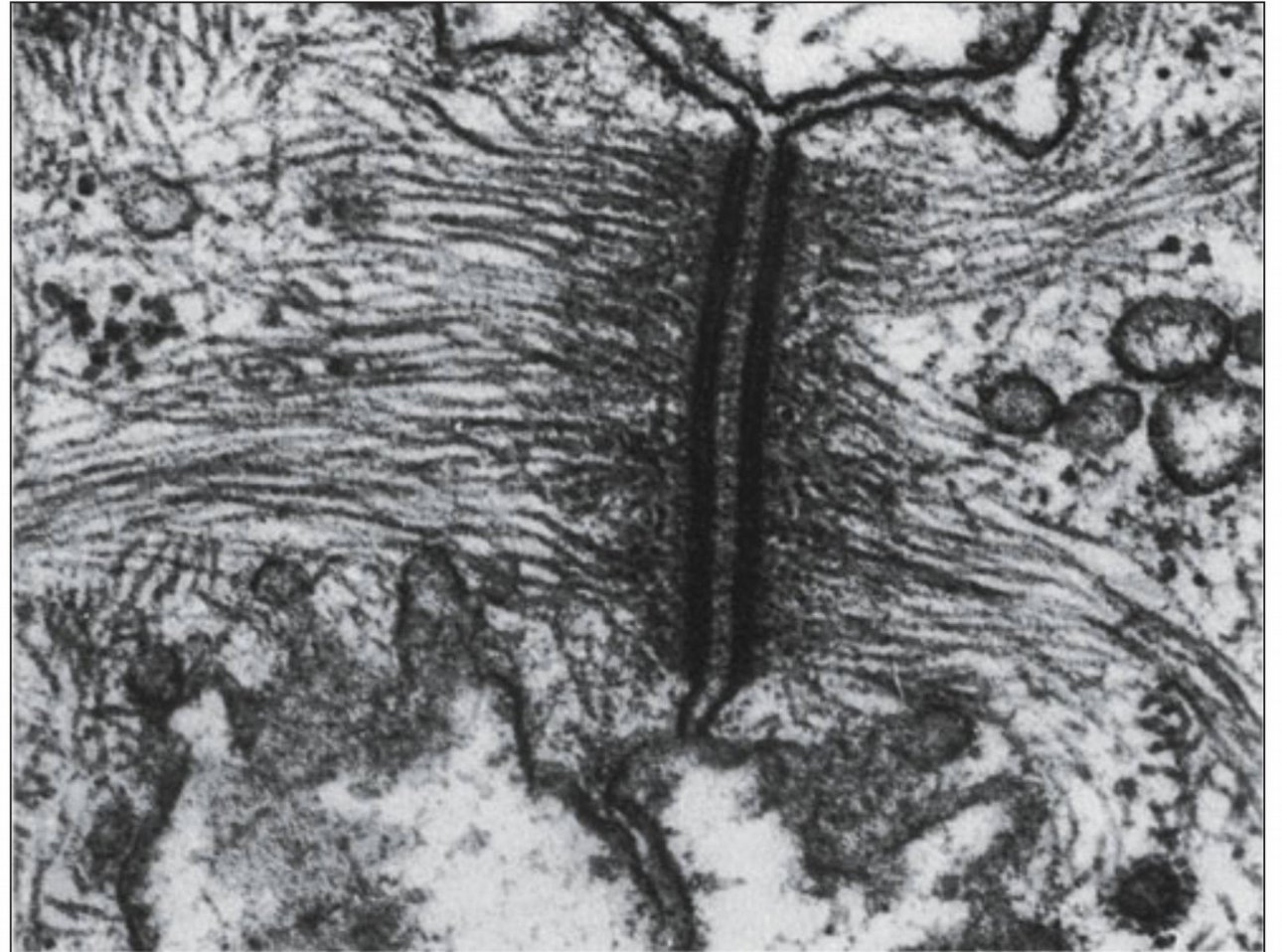
0.5 μm

Lumen

freeze-fractured
membrane

Jonctions : desmosomes

- Patches
- Cadhérine, protéines adaptatrices, kératine



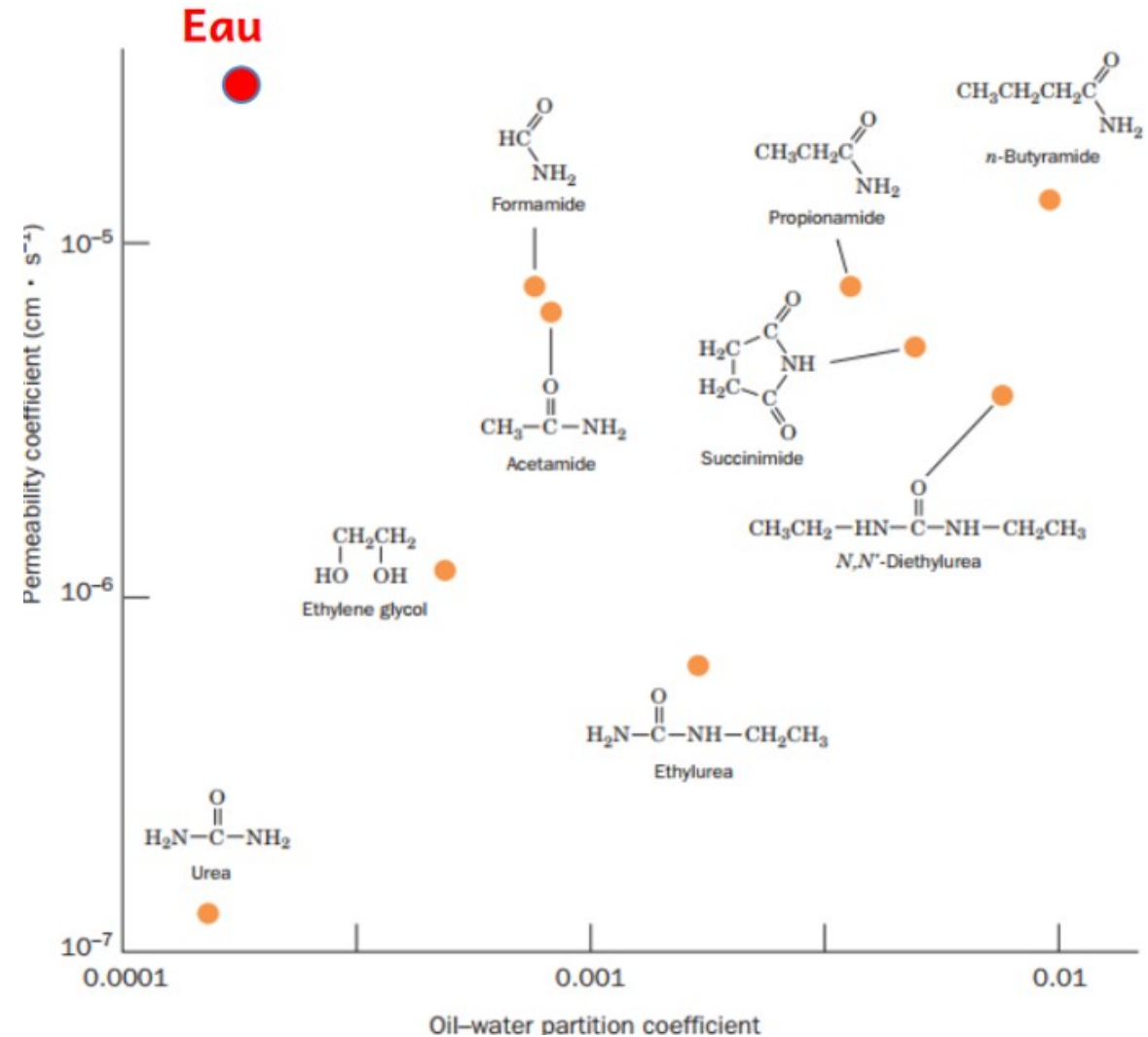
0.5 μm

Transports membranaires

- Potentiel électrochimique
- Potentiel chimique
- Potentiel électrique
- Sens de passage
- Exergonique VS endergonique

Transports membranaires

- Coefficient de partition
- Hydrophobe → diffusion simple facilitée



Transports membranaires : cas de l'eau

- Transport par des aquaporines

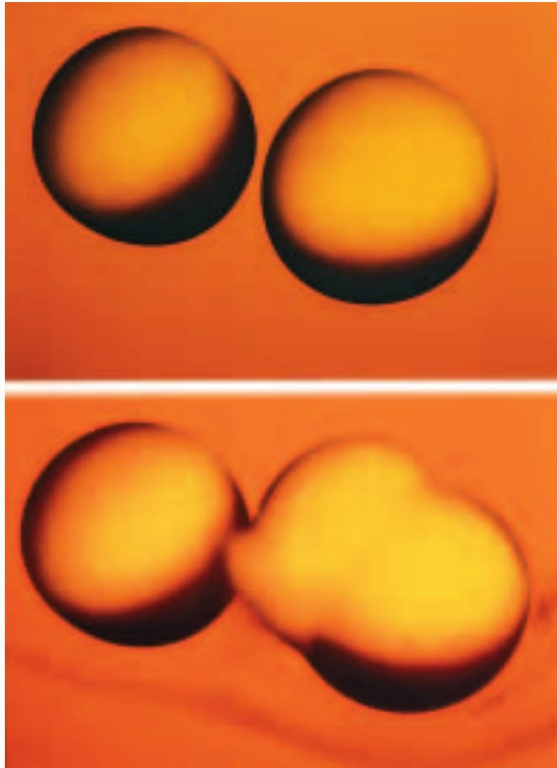
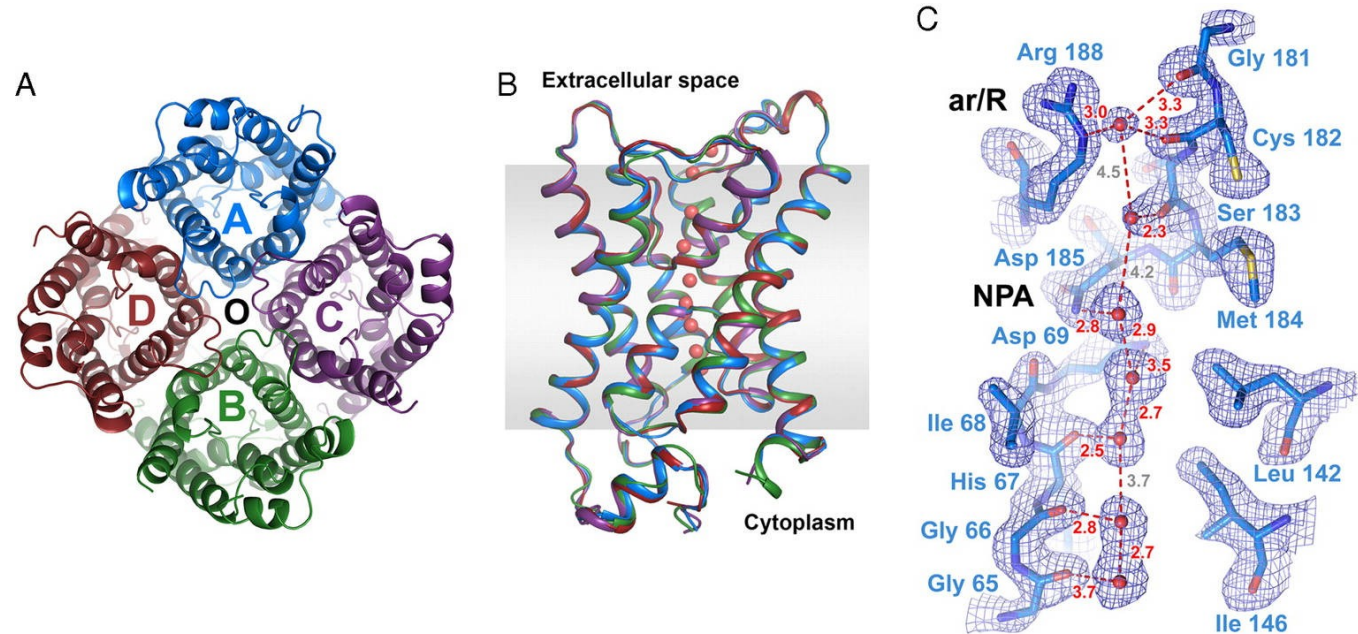
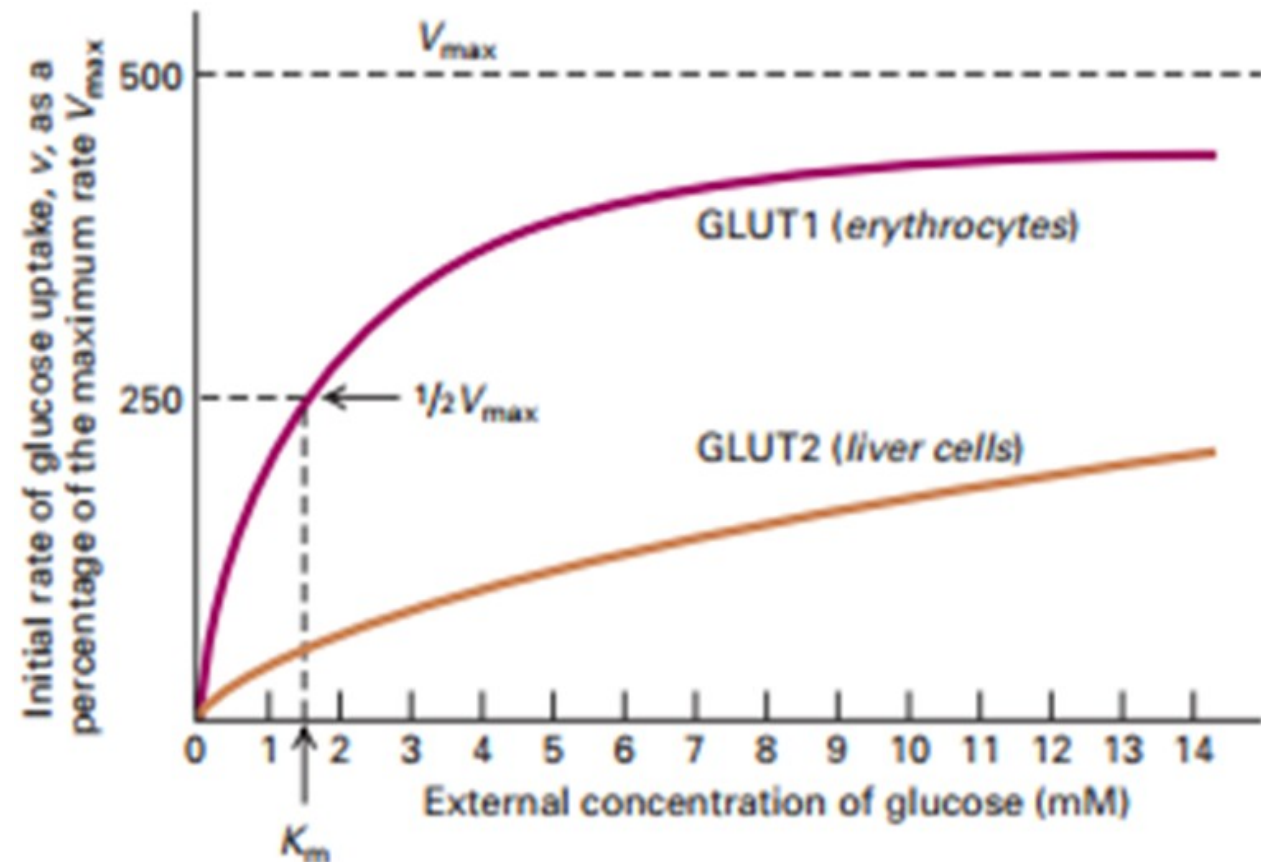


Figure 2. Functional expression of AQP1 water channels in *Xenopus laevis* oocytes. Control oocyte (left) was injected with water; AQP1 oocyte (right) was injected with cRNA. The oocytes were transferred to hypotonic buffer. After 30 seconds (top) the AQP1 oocyte has begun to swell; after 3 minutes (bottom), the AQP1 oocyte has exploded. Modified and reprinted from [Science](#) with permission (Preston *et al.*, 1992).



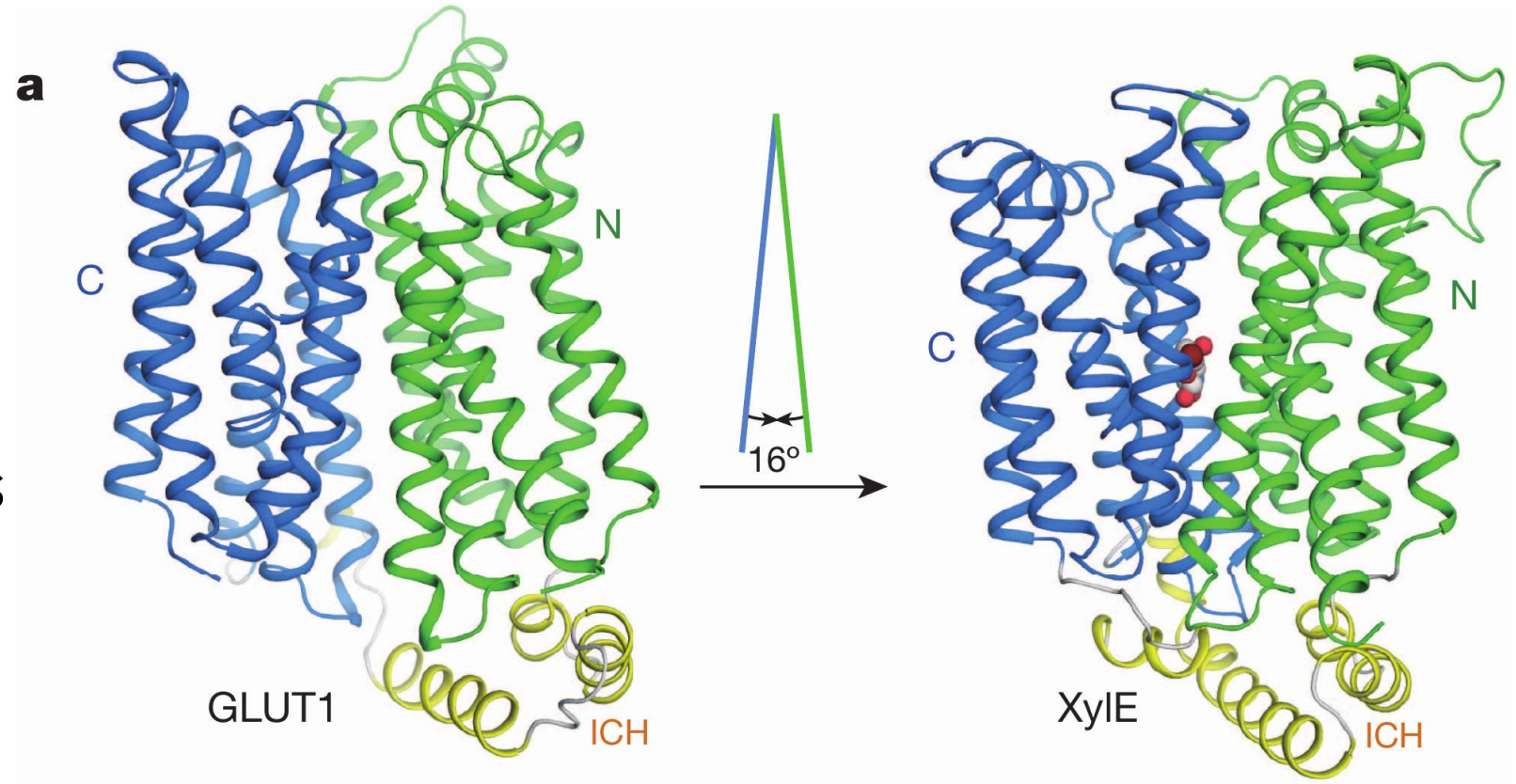
Transports membranaires : cas du glucose

- Transport par des transporteurs spécialisés (GLUT)
- Famille de transporteurs aux cinétiques variables



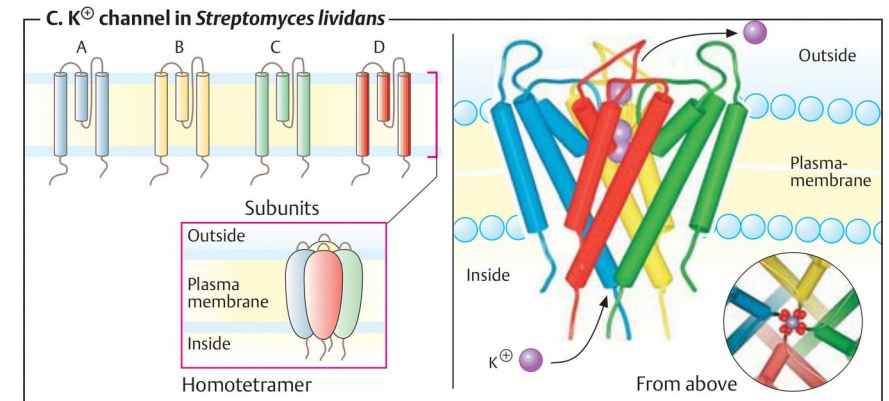
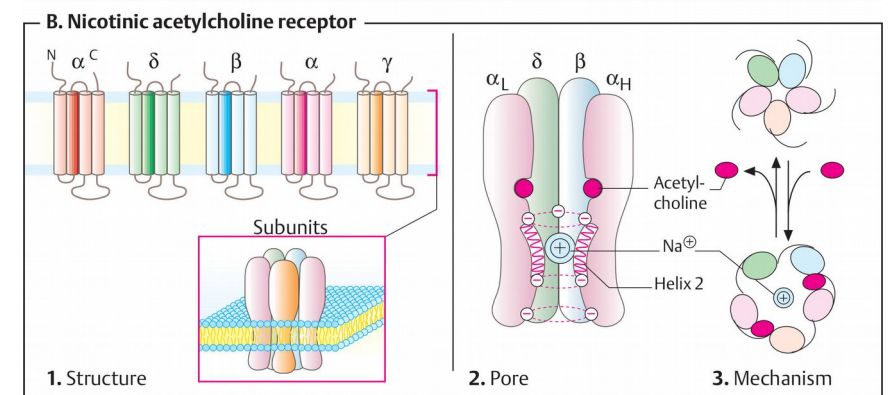
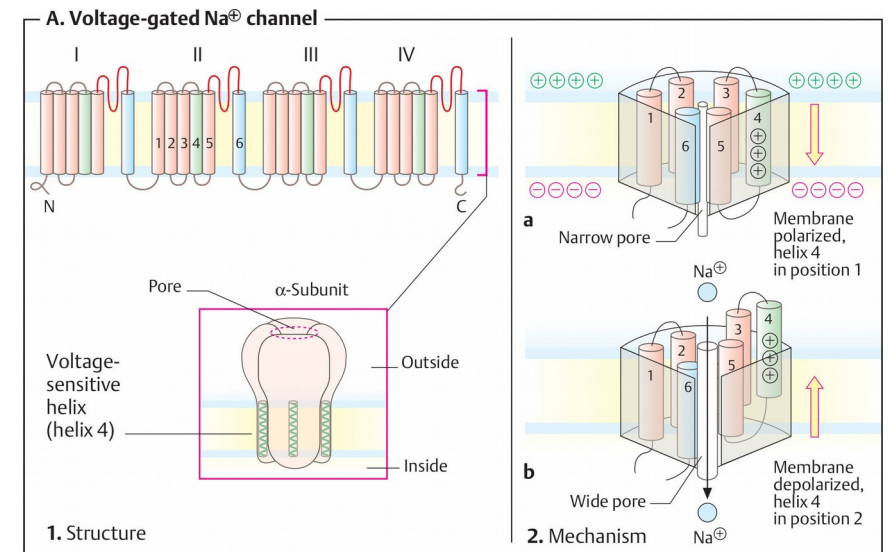
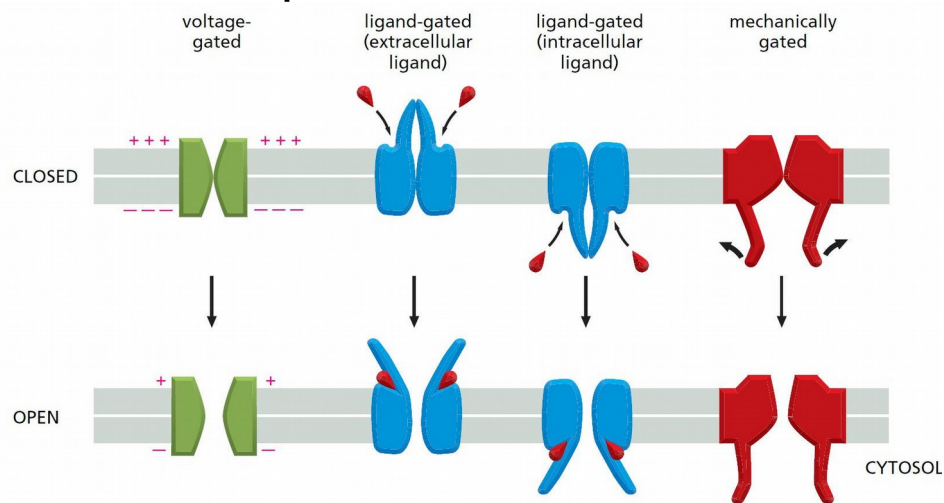
Transports membranaires : cas du glucose

- Transport par des transporteurs spécialisés (GLUT)
- Famille de transporteurs aux cinétiques variables



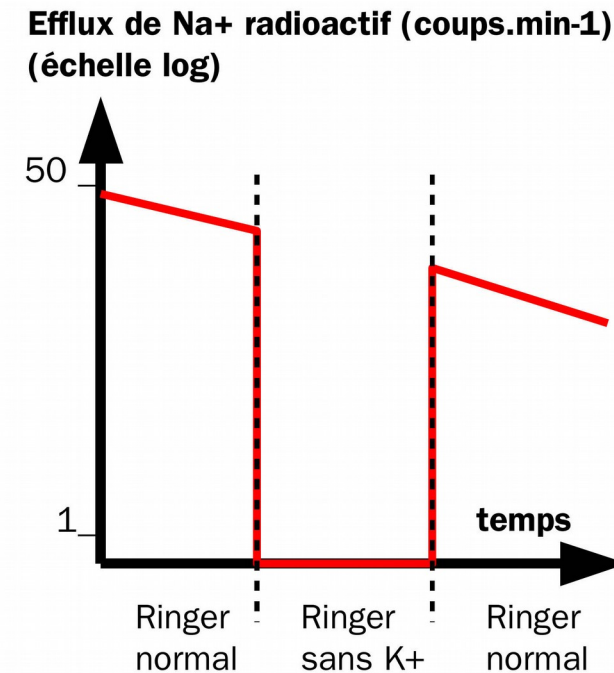
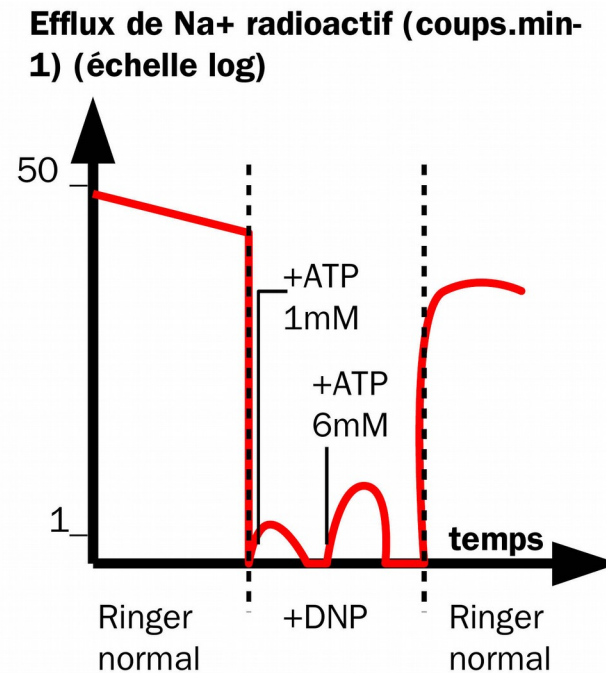
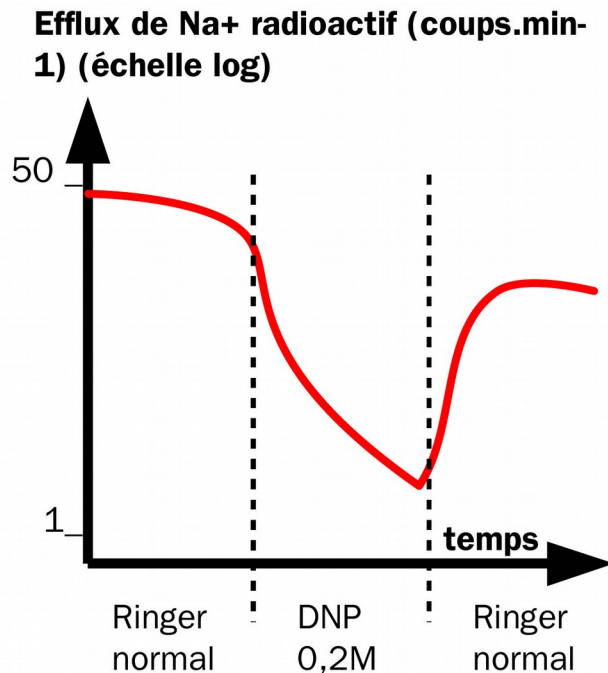
Transports membranaires : cas des ions

- Canaux ioniques
 - De fuite
 - Voltage-dépendants
 - Chimio-dépendants
 - Mécano-dépendants



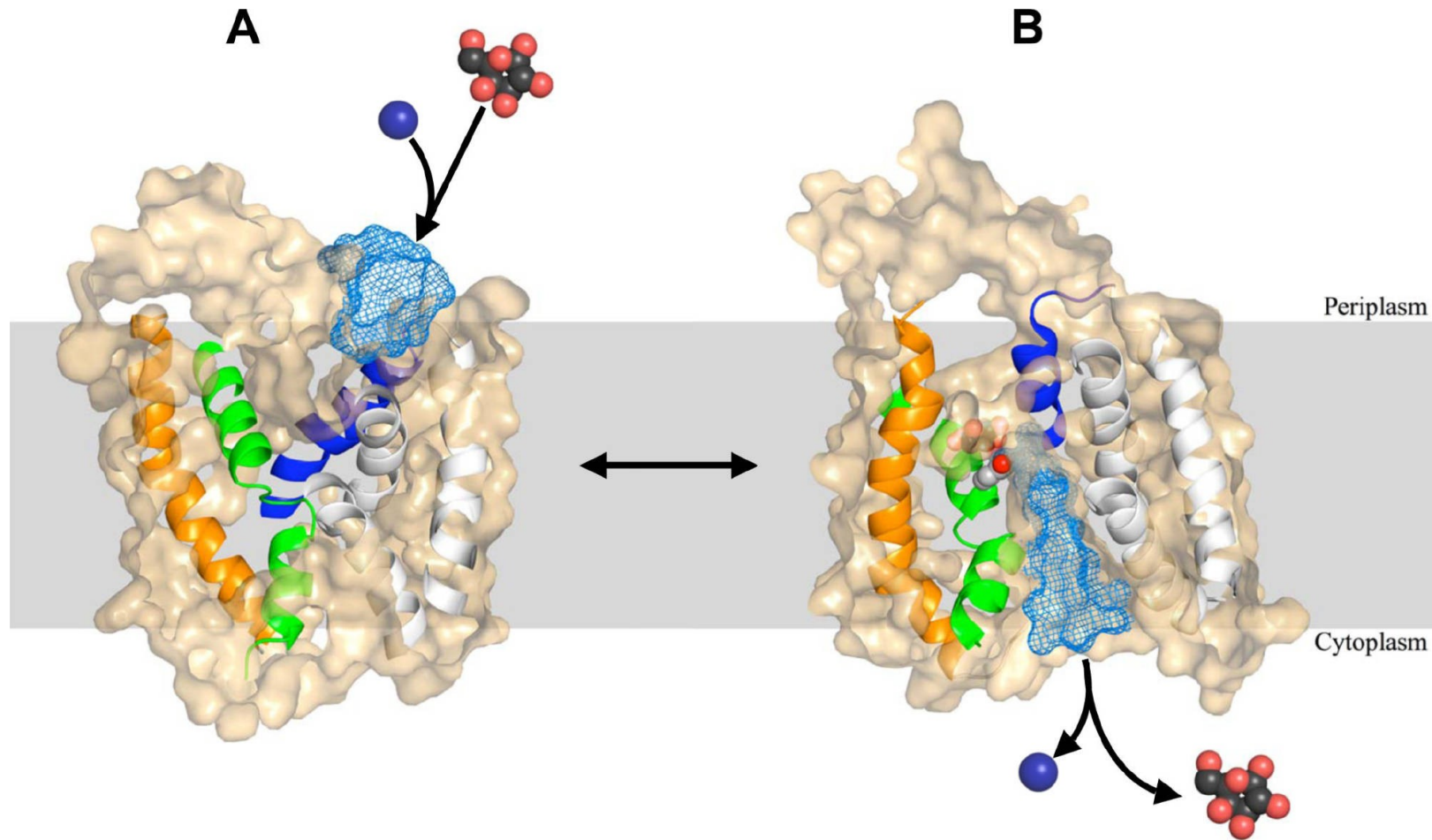
Transports membranaires : découverte des transports actifs

- Sur axone de Calmar
- Nobel 1963



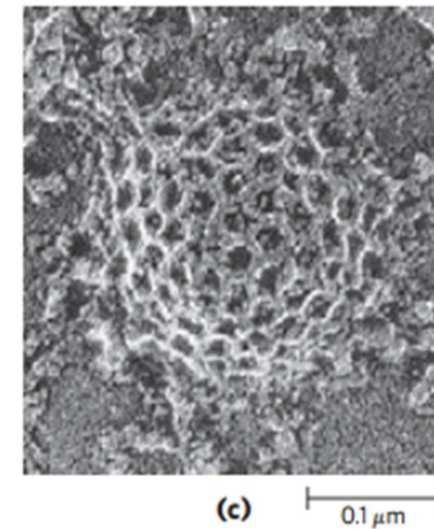
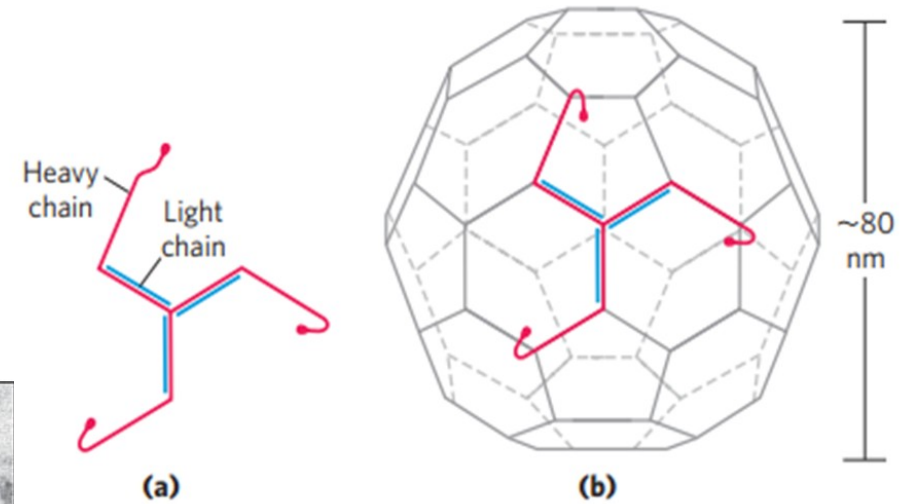
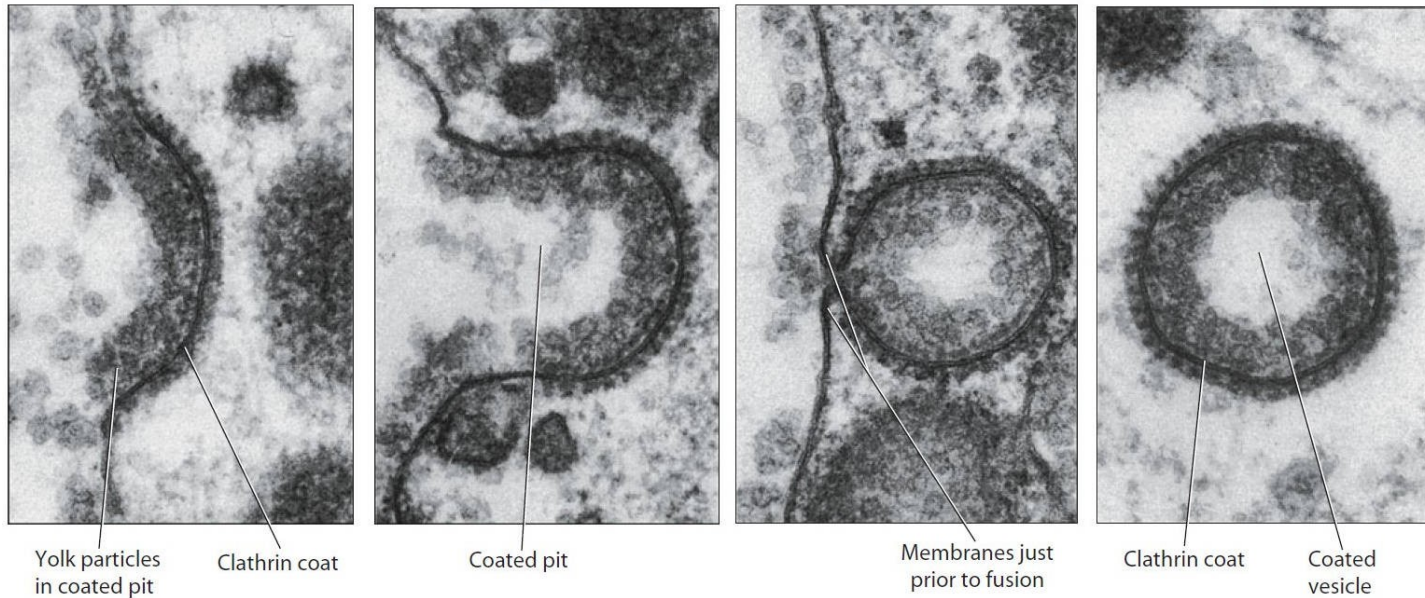
Transports membranaires : transports actifs secondaires

- SGLT



Transports membranaires : endocytose

- Molécules de clathrine



Transports membranaires : endocytose

- Modèle d'endocytose
- Intervention de la dynamine

